Original research article

Evaluation Of The Antimicrobial Susceptibility Pattern And The Pathogenesis Of Surgical Site Infection: An Observational Study

Dr. Deepti Kiran¹, Dr. Rajesh Prasad², Dr. Arjun Lal³

¹ Tutor, Department of Microbiology, Anugrah Narayan Magadh Medical College and Hospital, Gaya, Bihar, India.

² Assistant Professor, Department of Microbiology, Anugrah Narayan Magadh Medical College and Hospital, Gaya, Bihar, India.

³Associate Professor & HOD, Department of Microbiology, Anugrah Narayan Magadh Medical College and Hospital, Gaya, Bihar, India

Corresponding Author: Dr. Rajesh Prasad

Abstract

Aim: The aim of the present study to evaluate the antimicrobial susceptibility pattern of microorganisms involved in the pathogenesis of surgical site infection.

Material and methods: The study was a cross sectional study which was carried in the Department of Microbiology, Anugrah Narayan Magadh Medical College and Hospital, Gaya, Bihar, India for one year. Using sterile cotton swabs, two pus swabs/ wound swabs were collected aseptically from each patient suspected of having SSI. Gram stained preparations were made from one swab for provisional diagnosis. The other swab was inoculated on nutrient agar, 5% sheep blood agar (BA) and MacConkey agar (MA) plates and incubated at 37°C for 24-48 hours before being reported as sterile. Growth on culture plates was identified by its colony characters and the battery of standard biochemical tests. All the isolates were tested for antimicrobial susceptibility by Kirby Bauer disk diffusion technique on Muller Hinton Agar.

Results: Out of 420 samples, 220 samples were culture positive (52.38%). Among 220 positive samples 120 (54.55%) were males. Maximum no. of culture positive samples in age 20-30 years (32.73%) followed by 30-40 (17.73%) and then followed by 40-50 (14.09%) of age group respectively. Out of 220 culture positive samples *S.aureus* (27.27%) was the most common pathogen isolated followed by Escherichia coli. (22.73%), Citrobacter spp. (14.54%) and *Pseudomonas aeruginosa* (12.27%) respectively. Among gram negative bacilli, E.coli was most sensitive to Imipenem 90%) followed by Amikacin (78%) and Piperacillin Tazobactam (74%) whereas for Citrobacter spp., Imipenem (81.25%) followed by Gentamicin (53.13%), Ciprofloxacin (46.87%) was the drug of choice then for Klebsiella spp., Imipenem (76.19%) followed by Gentamicin (47.62%), Amikacin (47.62%) was the drug of choice. For Pseudomonas aeruginosa, Imipenem (62.96%) followed by Piperacillin Tazobactam (59.26%), Gentamicin (51.85%) was the drug of choice and for Enterobacter *spp.*, Imipenem (83.33%) followed by Amikacin (66.67%), Piperacillin Tazobactam (66.67%) showed maximum sensitivity (Table 4). Among gram positive organism, S.aureus showed maximum antibiotic sensitivity to Linezolid (93.33%) followed by Vancomycin (91.67%), Amikacin (81.67%) whereas CONS was sensitive to Linezolid (100%) followed by Vancomycin (93.75%), and Gentamicin (87.5%).

Conclusion: We conclude that the increasing resistance to antimicrobials increases the risk of morbidity and mortality; therefore there is urgent need of implementation of measures to restrict the health care associated infection. Rational use of antimicrobials, proper hygiene, and strict asepsis should be applied in all health care.

Volume 07, Issue 11, 2020

Introduction

Infections are encountered by all surgeons post operatively. Sometimes postoperative infections lead to death. Death from infection was so common after compound fractures, amputations and gunshot wounds. Now a day's surgical site infections are mainly due to hospital acquired infections and irrational use of antibiotics, surgical site infection (SSI) is one of postoperative complications in any surgery. Surgical site infections (SSI) are among most common nosocomial infections and are encountered in approximately 2%-5% of patients undergoing surgery. Infections which occur during the time of hospital stay and were not present or in incubating stage, during the time of hospital admission are considered as health care associated infections.¹ Health care associated infection possess major problem for both doctor as well as patients. Health care associated infection prolongs hospital stay which leads to financial burden to the patients. It has been reported that in United States of America the death frequency is about 88,000 every year despite of estimated cost of management of health care associated infections is about 4.5 billion dollar.² Among health care associated infections, surgical site infections are the second most common after Urinary tract infection.³ Surgical site infections (SSI), one of the most common causes of nosocomial infections are a common complication associated with surgery, with a reported incidence rates of 2-20%.⁴ They are responsible for increasing the treatment cost, length of hospital stay and significant morbidity and mortality. Despite the technical advances in infection control and surgical practices, SSI still continue to be a major problem, even in hospitals with most modern facilities.⁵ These infections are usually caused by exogenous and/ or endogenous micro organisms that enter the operative wound either during the surgery (primary infection) or after the surgery (secondary infection). Primary infections are usually more serious, appearing within five to seven days of surgery.⁶ Majority of SSIs are uncomplicated involving only skin and subcutaneous tissue but sometimes can progress to necrotizing infections. The usual presentation of infected surgical wound can be characterized by pain, tenderness, warmth, erythema, swelling and pus formation.^{7,8} A number of patient related factors (old age, nutritional status, pre existing infection, co-morbid illness) and procedure related factors (poor surgical technique, prolonged duration of surgery, pre operative part preparation, inadequate sterilization of surgical instruments) can influence the risk of SSIs significantly.⁵ In addition to these risk factors, the virulence and the invasiveness of the organism involved, physiological state of the wound tissue and the immunological integrity of the host are also the important factors that determine whether infection occurs or not.⁹

Surveillance data suggest that the types of causative organisms associated with SSI have not significantly changed over the past 10–15 years; however, the proportion of different types of causative organisms has changed. Antimicrobial-resistant organisms are causing an increasing proportion of SSIs, and there has been a rise in the number of infections caused by atypical bacterial and fungal organisms. These changing proportions have been attributed to the increasing acuity of surgical patients, the increase in the number of immunocompromised patients, and the increasing use of broad-spectrum antibiotics.¹⁰

Surgical site infections are the second most common cause of Nosocomial infections.¹¹ Surgical site infections are still a threat to patients, in spite of the newer antibiotics available today. Although properly administered antibiotics can reduce postoperative surgical site infections secondary to bacterial contamination, widespread use of prophylactic antibiotics can lead to emergence of multi drug resistant bacteria. The higher rates of surgical site infections are associated with higher morbidity, mortality and increased medical expenses.¹²

In developing countries like India, where hospitals have inadequate infrastructure, poor infection control practices, overcrowded wards and practice of irrational use of antimicrobials, the problem of SSIs gets more convoluted. The aim of the present study to

evaluate the antimicrobial susceptibility pattern of microorganisms involved in the pathogenesis of surgical site infection.

Material and methods

The study was a cross sectional study which was carried in the Department of Microbiology, Anugrah Narayan Magadh Medical College and Hospital, Gaya, Bihar, India for one year, after taking the approval of the protocol review committee and institutional ethics committee. Total 420 patients with SSIs either sex or any age, who had surgical wound pus, discharge, or signs of sepsis were include in this study. Patients with cellulitis and suture abscess were excluded from this study.

Using sterile cotton swabs, two pus swabs/ wound swabs were collected aseptically from each patient suspected of having SSI. Gram stained preparations were made from one swab for provisional diagnosis. The other swab was inoculated on nutrient agar, 5% sheep blood agar (BA) and MacConkey agar (MA) plates and incubated at 37°C for 24-48 hours before being reported as sterile. Growth on culture plates was identified by its colony characters and the battery of standard biochemical tests.^{13,14} All the isolates were tested for antimicrobial susceptibility by Kirby Bauer disk diffusion technique on Muller Hinton Agar and results were interpreted in accordance with Clinical Laboratory Standards Institute guidelines.¹⁵ Antibiotics used for susceptibility testing were: Amikacin, Ampicillin / Sulbactam, Ceftriaxone, Ciprofloxacin, Gentamicin, Piperacillin-Tazobactum, Imipenem, Azithromycin, Vancomycin, Linezolid, Ofloxacin, Cefoxitin.

Statistical Analysis: Data was entered in Microsoft excel spreadsheet and analysed using appropriate statistical software application.

Results

Out of 420 samples, 220 samples were culture positive (52.38%) (Table 1). Among 220 positive samples 120 (54.55%) were males (Table 1). The age wise distribution of the gender has been shown in the (Table 2) with maximum no. of culture positive samples in age 20-30 years (32.73%) followed by 30-40 (17.73 %) and then followed by 40-50 (14.09%) of age group respectively. Out of 220 culture positive samples S.aureus (27.27%) was the most common pathogen isolated followed by *Escherichia coli*. (22.73%), *Citrobacter spp*. (14.54%) and Pseudomonas aeruginosa (12.27%) respectively (Table 3). Among gram negative bacilli, *E.coli* was most sensitive to Imipenem 90%) followed by Amikacin (78%) and Piperacillin Tazobactam (74%) whereas for *Citrobacter spp.*, Imipenem (81.25%) followed by Gentamicin (53.13%), Ciprofloxacin (46.87%) was the drug of choice then for Klebsiella spp., Imipenem (76.19%) followed by Gentamicin (47.62%), Amikacin (47.62%) was the drug of choice. For Pseudomonas aeruginosa, Imipenem (62.96%) followed by Piperacillin Tazobactam (59.26%), Gentamicin (51.85%) was the drug of choice and for Enterobacter spp., Imipenem (83.33%) followed by Amikacin (66.67%), Piperacillin Tazobactam (66.67%) showed maximum sensitivity (Table 4). Among gram positive organism, S.aureus showed maximum antibiotic sensitivity to Linezolid (93.33%) followed by Vancomycin (91.67%), Amikacin (81.67%) whereas CONS was sensitive to Linezolid (100%) followed by Vancomycin (93.75%), and Gentamicin (87.5%) (Table 5).

Table 1: Gender wise d	istribution of Culture	positive Patients

Gender	No of patients=220
Male	120 (54.55%)
Female	100(45.45%)

Volume 07, Issue 11, 2020

Age in year	Culture Positive
Below 20	30 (13.63)
20-30	72 (32.73)
30-40	39(17.73)
40-50	31 (14.09)
50-60	27(12.27)
Above 60	21 (9.55)

Table 2: Age wise Distribution of Culture Positive Patients

Table 3: Distribution of Organisms Causing Surgical Site Infection

Organism	No. of isolates (%)
Staphylococcus aureus	60(27.27)
Escherichia coli	50 (22.73)
Citrobacter spp.	32(14.54)
Pseudomonas aeruginosa	27(12.27)
Klebsiella spp.	21 (9.55)
CONS	16 (7.27)
Enterobacter spp.	12(5.45)
Acinetobacter spp.	1 (0.4)
Proteus spp.	1(0.4)
Total	220

Table 4: In-Vitro Antibiotic Sensitivity in Isolated Gram Negative Bacteria

Drugs	Escherichia	Citrobacter	Klebsiella	Pseudomonas	Enterobacter
	coli	spp.	<i>spp</i> . (%)aeruginosa (%)spp. (%)
	(%)(n=50)	(%) (n=32)	(n=21)	(n=27)	(n=12)
	S	S	S	S	S
Gentamicin	33 (66)	17(53.13)	10 (47.62)	14 (51.85)	5(41.67)
Ciprofloxacin	15. (30)	15 (46.87)	8(38.09)	14(51.85)	5(41.67)
Piperacillin/	37 (74)	12 (37.5)	7 (33.33)	16 (59.26)	8(66.67)
Tazobactam					
Amikacin	39 (78)	15 (46.87)	10 (47.62)	15 (55.55)	8(66.67)
Ampicillin/	18 (36)	9(28.12)	6 (28.57)	9 (33.33)	3 (25)
Sulbactam					
Impinem	45(90)	26 (81.25)	16 (76.19)	17 (62.96)	10(83.33)
Ceftriaxone	14 (28)	10 (31.25)	5 (23.81)	12 (44.44)	2 (16.67)

Table 5: In-Vitro Antibiotic Sensitivity in Isolated Gram Positive Bacteria

Drugs	Staphylococcus aureus (%)	CONS (%) (n=16)	
	(n=60)		
	S	S	
Azithromycin	37(61.67)	10 (62.5)	
Vancomycin	55 (91.67)	15(93.75)	
Linezolid	56(93.33)	16 (100)	
Gentamicin	47 (78.33)	14 (87.5)	
Ofloxacin	48 (80)	12 (75)	
Cefoxitin	41 (68.83)	9 (56.25)	
Amikacin	49 (81.67)	11(68.75)	

Volume 07, Issue 11, 2020

Discussion

Despite the advances in surgical techniques and better understanding of the pathogenesis of wound infection, management of SSIs remains a significant concern for surgeons and physicians in a health care facility. Patients with SSIs face additional exposure to microbial populations circulating in a hospital set up which is always charged with microbial pathogens. The unrestrained and rapidly spreading resistance to the available array of antimicrobials further contributes to the existing problem. Most of the SSIs are hospital acquired and vary from hospital to hospital.

In the present study the Culture positive SSI rate was 52.38%. Whereas various other studies from India have shown the rate of SSI to vary from 6.1% to 38.7%.¹⁶⁻¹⁹ The main Reason behind may be due to the lack of attention towards the infection control measures, inappropriate hand hygiene practices and overcrowded hospitals. In our study, it was observed that rate of infection was higher in male patients (54.55%). The results were similar to a study by Vikrant Negi et al, who reported that (74.6%) males were more commonly affected than females (25.5%).²⁰ In contrast to our study Gangania P et al reveals that 20% Females shows almost equal distribution of 19% of males.²¹

The findings in the study revealed that maximum no. of culture positive samples in age 20-30 years (32.73%) followed by 30-40 (17.73%) and then followed by 40-50 (14.09%) of age group respectively. Similar results was showed by Pooja Singh Gangania who concluded that maximum no of SSI was in 16-45 years of age group (24%) patient. This may be due to heavy work load, stress at this age group and less number of patients.²¹ *S.aureus* (27.27%) was the most common pathogen isolated followed by *E.coli* (22.73%). This result is consistent with reports from other studies SP Lilani, Mulu W.^{17,22} *S. aureus* infection is most likely associated with endogenous source as it is a member of the skin and nasal flora and also with contamination from environment, surgical instruments or from hands of health care workers.²⁰

In the present study among gram negative bacilli, *E.coli* was most sensitive to Imipenem 90%) followed by Amikacin (78%) and Piperacillin Tazobactam (74%). The findings are consistent with the previous study conducted by M. saleem et al who also showed that *E. coli* showed high sensitivity to Imipenem.²³ In this study *Citrobacter spp.*, Imipenem (81.25%) followed by Gentamicin (53.13%), Ciprofloxacin (46.87%) was the drug of choice then for *Klebsiella spp.*, Imipenem (76.19%) followed by Gentamicin (47.62%), Amikacin (47.62%) was the drug of choice. The findings are consistent with the study conducted by Jyoti Sonawane et al who also showed that *Citrobacter* and *Klebsiella* showed high sensitivity to Imipenem.²⁴

Pseudomonas aeruginosa, Imipenem (62.96%) followed by Piperacillin Tazobactam (59.26%), Gentamicin (51.85%) was the drug of choice. Similar results were shown by Jyoti Sonawane et al.²⁴ Imipenem, Piperacillin/ Tazobactum, Gentamicin and Amikacin were found to be more efficient antibiotics against gram negative bacilli . Similar results were observed by M. saleem et alwho showed that Amikacin, Imipenem, Piperacillin/ Tazobactum, were found to be more efficient antibiotics against gram negative bacilli.²³ Among gram positive organism, *S.aureus* showed maximum antibiotic sensitivity to Linezolid (93.33%) followed by Vancomycin (91.67%), Amikacin (81.67%). This was in consistent with the study by Prem Prakash Singh et al., 2015 who also concluded that *S. aureus* was sensitive to Vancomycin (100%), Linezolid (100%).²⁵ Linezolid and Vancomycin were found to be more efficient antibiotics against gram positive cocci . This finding was in tandem with the study conducted by Vikrant Negi et al., 2015, who also reported that Vancomycin and Linezolid found to be more efficient antibiotics against gram positive cocci.²⁰

Conclusion

We conclude that the increasing resistance to antimicrobials increases the risk of morbidity and mortality; therefore there is urgent need of implementation of measures to restrict the health care associated infection. Rational use of antimicrobials, proper hygiene, and strict asepsis should be applied in all health care.

References

- 1. Ananthanarayan R, Panikar CJK, editors. Textbook of microbiology, 5th ed., Chennai: Orient Longman, 2005.
- 2. Weinstein RA, Infection control in the hospital. In: Braunwald E, Fauci AS, Kasper DL, Harrison's principles of internal medicine. 15th ed., New York: McGraw-Hill, 2001; 1; 853-856.
- **3.** Anvikar RA, et.al A one year prospective study of 3280 surgical wounds. I.J.M.M. 1999; 17 (3): 129-32
- 4. Hohmann C, Eickhoff C, Radziwill R, Schulz M. Adherence to guidelines for antibiotic prophylaxis in surgery patients in German hospitals: a multicentre evaluation involving pharmacy interns. Infection. 2012;40(2):131-37.
- 5. Owens CD, Stoessel K. Surgical site infections: epidemiology, microbiology and prevention. J Hosp Infect. 2008;70(Suppl 2): 3-10.
- 6. Pradhan GB, Agrawal J. Comparative study of post operative wound infection following emergency lower segment caesarean section with and without the topical use offusidic acid. Nepal Med Coll J. 2009;11(3):189-91.
- Ahmed MI. Prevalence of nosocomial wound infection among postoperative patients and antibiotics patterns at teaching hospital in Sudan. N Am J Med Sci .2012;4(1):29-34.
- 8. Mulu W, Kibru G, Beyene G, Datie M. Postoperative nosocomial infections and antimicrobial resistance patterns of bacterial isolates among patients admitted at FelegeHiwot Referral Hospital, Bahirdar, Ethiopia. Ethiop J Health Sci. 2012;22(1):7-18.
- 9. Masaadeh HA, Jaran AS. Incident of Pseudomonas aeruginosa in post-operative wound infection. Am J Infect Dis. 2009;5:1–6.
- SievertDM, RicksP, EdwardsJR, SchneiderA, PatelJ, SrinivasanA, et al. Antimicrobial-resistant pathogens associated with healthcare-associated infections: Summary of data reported to the national healthcare safety network at the centers for disease control and prevention, 2009-2010. Infect Control Hosp Epidemiol 2013;34:1-4.
- 11. Sohil Ahmed Khan, Padma G.M .Rao, Anand Rao, Gabriel Rodrigues. Survey and evaluation of antibiotic prophylaxis usage in surgery wards of tertiary level institution before and after the implementation of clinical guidelines. Indian Journal of Surgery 2006; vol. 68 (3): 150-156.
- 12. Sasse A, Mertens R, and Sion JP, et al. Surgical prophylaxis in Belgian hospitals Estimate of costs and potential savings. J Antimicrobial Chemotherapy 1998; 41: 267-272
- 13. MacFaddin J. Biochemical Tests for Identification of Medical Bacteria. 3 rded. Philadelphia: Lippincott Williams and Wilkins; 1976.
- 14. Forbes BA, Sahm DF, Weissfeld AS. Bailey and Scott's Diagnostic Microbiology.10th ed. St. Louis, Misssouri, USA: Mosby Inc.; 1998

- 15. Clinical and Laboratory Standard Institute. Performance Standards for Antimicrobial Susceptibility Testing.2007;1(1).M2 A9. Pennsylvania, USA: Clinical and Laboratory Standard Institute.
- 16. Malik S, Gupta A, Singh PK, Agarwal J, Singh M. Antibiogram of aerobic bacterial isolates from postoperative wound infections at a tertiary care hospital in india. Journal of Infectious Diseases Antimicrobial Agents. 2011;28:45-51.
- 17. Lilani SP, Jangale N, Chowdhary A, Daver GB. Surgical site infection in clean and clean-contaminated cases. Indian J Med Microbiol. 2005;23(4):249-52.
- 18. Khan A K A, Rashed MR, Banu G. A Study on the Usage Pattern of Antimicrobial Agents for the Prevention of Surgical Site Infections (SSIs) in a Tertiary Care Teaching Hospital. J Clin Diagn Res. 2013 ;7(4):671-4.
- 19. Chakarborty SP, Mahapatra SK, Bal M, Roy S. Isolation and identification of [14] vancomycin resistant Staphylococcus aureus from postoperative pus sample. Al Ameen J Med Sci. 2011; 4(2):152-68.
- 20. Negi V, Pal S Juyal D, Sharma M K, Sharma N. Bacteriological Profile of Surgical Site Infections and Their Antibiogram: A Study From Resource Constrained Rural Setting of Uttarakhand State, India. Journal of Clinical and Diagnostic Research. 2015;9(10)
- 21. Gangania P S, Singh V A, Ghimire S S. Bacterial Isolation and Their Antibiotic Susceptibility Pattern from Post-Operative Wound Infected Patients. Indian J Microbiol Res 2015; 2(4):231-235.
- 22. Mulu W, Kibru G, Beyene G, Damtie M. Postoperative nosocomial infections and antimicrobial resistance pattern of bacteria isolates among patients admitted at Felege Hiwot Referral Hospital, Bahirdar, Ethiopia. Ethiopian Journal of Health Sciences. 2012; 22(1):7–18.
- 23. Saleem M, Subha TV, Balamurugan R, Kaviraj M, Gopal R. Bacterial Profile and Antimicrobial Susceptibility Pattern of Surgical Site Infections A Retrospective Study. Indian Journal Of Applied .2015;5(10):ISSN 2249-555X.
- 24. Sonawane J, Kamath N, Swaminathan R, Dosani K. Bacteriological profile of Surgical Site Infections and their Antibiograms in A Tertiary Care Hospital Navi Mumbai. Bombay Hospital Journal.2010;52(3).
- 25. Singh PP, Begum R, Singh S, Singh MK. Identification and Antibiogram of the Microorganisms Isolated from the Post operative Surgical Site Infections among the patients admitted in the hospital TMMC & RC, Moradabad. European journal of biomedical and pharmaceutical sciences. 2015;2(4):932-942.

Received: 12-09-2020 || Revised: 02-10-2020 || Accepted: 25-10-2020