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DETERMINATION OF THE NUMBER OF BIOLOGICALLY ACTIVE SUBSTANCES PREPARING THE MEDICINE PROPERTIES OF TREES FOR CORRECTION OF ANIMAL POISONED PESTICIDES

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Abstract: ***The total amount of biologically active compounds - antioxidants, flavonoids, and vitamin C in different trees for example in Morusnigra L., Prunuspersica L., Serasus Vulgaris L., and Juglansregia L. leaves were found at different seasons. The results showed that the highest concentration of antioxidants in spring was found in walnuts and peaches than in the summer season. The number of walnut leaves was about 9.105 ± 0.05 mg/g, and in peach, leaves were about 9.082 ± 0.075 mg/g. In spring, the content of flavonoids in the leaves of grapes was high and amounted to 4721 ± 0.521 mg/g, in the summer their number was 3825 ± 0.255 mg/g, and in the fall there was a decrease in flavanoids to 1.460 ± 0.2253 mg/g. In the spring, in the stalks of grapes, flavonoids were 4.386 ± 0.046 mg/g, in cocoons 0.348 ± 0.023 mg/g.***

Keywords: antioxidant, flavonoids, Morusnigra L., Prunuspersica L., Serasus vulgaris L., Juglansregia L., Vitisvinifera, extract, leaf, grapes.

Introduction It is known that agricultural pesticides have adverse effects on the environment and have high biological activity, adversely affects human and animal organs, and leads to liver transformation. Many xenobiotics break down the functions of liver cells. As a result, cell organelles - mitochondria and microsomes - are highly variable. These changes, in turn, cause injury to the entire organism. Our research has shown that 1/10 LD_{50} dose of carate pesticides sent by peroxide to the rats has been found to slowly release the organism within 40-50 days [1,2].

Our results show that after 10 and 20 days of poisoning, residual pesticides in animals' liver were 12.71% and 6.78% respectively. In the 30th and 40th days, 5.13% and 1.62% respectively. The pesticide poisonous rats had not been detected for 50 days when the pesticides were not chromatized. The results show that karate leads to an infringement of

several metabolism in the body and slowly leaves the body. At the same time, pesticides can change the structure of the membrane.

Also, there is an increase in perforated oxidation of lipids and changes in the number of fermental activity in liver mitochondria and microsomes under the influence of karate [2]. The most ingestion of enzymes was observed one day after the poisoning. After the poisoning, we observed that the rats that were injected with antioxidant herbs were restored to the liver mitochondria enzymes (citochrome-s-oxidase, Mg + 2-ATFase)[3]. We have found that several enzymes have changed in the liver of pregnant rats embryos[4, 5].

In our research, we have been searching for drugs that are cheaper and more affordable for correction, with antioxidant properties from local herbs. 5% of the leaves of herbs, which have the medicinal properties, have been extracted from the herbal extracts and 30 minutes after the poisoning, 4 days after 1 ml of extracts of medicinal herbs.

When the herbal leaf extract was injected with pesticide rats, it was observed that peroxidation of lipids was close to normal and that the activity of the enzymes was almost restored. On the 50th day of the poisoning, peroxidation of lipids and activation of membranic enzymes have been restored[1, 2]. This may allow the conclusion that it can be used to correlate the extract of medicinal herbs used in the research.

The main purpose of our scientific work: We detected the amount of vitamin C, total antioxidants, flavonoids, biologically active substances that demonstrate the antioxidant properties of extracts of walnuts, peaches and Morusalba leaf medicinal plants used for correction of pesticides poisoning.

Walnut is a healing plant and most important among fruit trees. The fruit of nuts increases the power of the mind and the mind by giving them strength. In the walnut fruit juice, there are 60-77% fat, protein, carbohydrates, vitamins C, E, K, B and P, carotene, iron, cobalt salts and other substances. The greenish crustacean of the ripe fruit is particularly rich in vitamin C. It also contains vitamins B_1, B_2, carotene, hydroquinone compounds, and up to 25% digesters. In the walnut leaf there are also various biologically active substances. It contains 4-5% vitamin C, vitamin B_1 and P, a large amount of carotene, inositol, hydroblyum, essential oils, flavonoids, ell age and gall ate acids, dyes, acids and compounds. The nut leaves of the walnut, nuts and fruit peel, have anti-inflammatory, bactericidal and fungicidal properties [6, 8,16,24].

Peach seeds contain fat, essential oil, amidaline glycoside, emulsion enzymes and other substances. The soft side of fruit contains carotenoids, sugars, vitamins B_1, B_2, organic acids, essential oils, pectin, potassium and magnesium salts, microelements. Seeds of medicine are obtained from oil. The leaf is used as a rose. Fertility is generally used as a supportive agent in heart disease [6, 8,16,24].

The fruits of mulberry have a blood-repellent nature; improves metabolism, can be used as a lightweight lubricant. Bone healing on the hooks, treatment of wounds can be used to raise the leaf's temperature. Bark from the horns can be used to treat the wounds when the temperature rises [6,13].

Materials and methods

Determination of the amount of vitamin C is based on the reduction of 2,6-dichlorophenolindophenol. The total amount of antioxidants was determined by Rogojin method [11]. The method is based on antioxidants oxidation of iron-III-chloride. In this

process iron-III-chloride to iron-II-chloride is returned, its content is determined by the intensity of the color formed by addition of 0-phenanthroline.

Detection of biologically active substances - flavonoids was conducted at the Laboratory of Laboratory of Medicines Standardization of the Tashkent Pharmaceutical Institute [7,25].

Getting the extract. In order to extract extracts of leaf 3,0 kg to the reactor and separated by 1:5 ratio (70% ethyl alcohol) were removed and separated by bismaterials. The separation vesselwas held for 3 hours at 40-50^{0}C. The hot discharge was poured and the process was repeated twice. The separator was cooled to room temperature and filtered, and the ethyl alcohol contained in the separation was discharged. The resulting cuttings were combined and dried at 45 ± 5 ° C on Spray Dryer TP-S15 spray dryer. As a result, a delicate, rapidly moisturizing extract was obtained in 19% of brown, odorless, fragrant and fragrant extracts [8, 9].

The mulberry leaf was separated from the lynx. Dried medicinal herbal products were stored in paper bags, in dark and dry places, at room temperature and removed for analysis.

Walnut leaf was harvested in May. In the walnut, in May, unripe fruits were harvested and 70% alcohol solution was prepared.

Research objects were medicinal plants meeting the requirements of the relevant regulatory documents and permitted for use in medical practice and are presented in the table below (Table 1) [10].

Table 1

Medicinal herbs selected for correction

T/p	Plants name, family	Used part	Impacting agents	It should be used in medicine
1.	Trifolium L. pratense, Buters Fabaceace Juglandaceae – Walnuts	leaf	Vitamins, Flavonides, vitamin C, excipients	Anti-inflammatory, bleeding. Tuberculosis in correcting pneumonia
2	Greek walnuts, walnuts, Juglans regia L.	Not matured	Vitamins, Flavonides, vitamin C, excipients, hydrolyglon	Vomiting, renal failure, liver, arteriosclerosis, chronic intestinal tract, low blood pressure
3.	Shoot, king-hold - Morus nigra L Tutors - Moraceae.	leaf	Polysaccharides, flavonoids, acyclic acids, excipients	Hypoglycemic, cholesterol - lowering, calm
3.	Category Vitis L. – Grape V. vinifera L. Vitaceae – Tokolders –	leaf	Essential oils Flavonoids Organic Acids	It is calming to improve its traditions
	Persica Mill. – Peaches Tour P. vulgaris Mill. – Peaches	fruits	Carotenoids, sugars, vitamins S, V1, V2, organic acids	Heart Disease Supporting Instrument

Chemical and commodity analysis of plant collections

XI - Examined the chemical composition and commodity analysis of plant specimens prepared according to the requirements of the State Pharmacopoeia [14].

At the same time, the most commonly accepted quality reactions and quantitative analysis methods and the relevant regulatory requirements were examined (Table 2) [7,14,15,26].

Table 2: **Results of chemical and commodity analysis of raw medicinal raw materials included in raw materials**

Raw materials / Indicators	Walnut leaf	Toe leaf	Morus albaleaf	Peach barley
Humidity,%	13.2	9.3	9.0	12.8
Totalprofit,%	7.6	3.98	7.0	9.2
10% solution that does not dissolve in chloride acid	0.48	2.06	3.0	1.1
Organiccompounds	0.4	1.4	0.4	5.81
Mineralcompounds	0.3	1.2	0.5	0.3
Groats 7 mm,%,	3	4.0	3	3
0.5 mmelongatedthinsection,%	1	1.5	2	2
Other regions of the plant (leaf, foam),%	1.8	3.3	0.5	0.4
Amount of the active substance	Flavonoids 9.4	Additives 15%	Polisaxarid 10%	Polsaxaridinulin 8%

Determination of the integrity of the raw materials was conducted in the following way[12, 17-20]:

1. 1 g crushed raw material was put into a 50 ml conical tube. Over 20 ml of 50% alcohol was added at a temperature of 60°C for 15 minutes, was heated in a water bath using a refrigerant. Boiling was cooled to room temperature and the white paper was filtered. The filter was dropped by 1ml. 1 ml of 96% alcohol, 0.1 g of magnesium powder and 1 ml of concentrated chloride acid were added to the remaining solution. It has been observed that there is a slightly red color, which is typically flavonoid.

2. Reactions to flavonoids (synod reactions)

- After 2 ml of solvent dissolved in a porcelain bottle, 2-3 ml of ethyl alcohol was dissolved in a water bath and 3-4 drops of concentrated hydrochloric acid was added and heated above magnesium metal and heated in a little water bath. The solution was red color.

- Add 2 ml of alcohol separation into the solution and add 5 drops of 1% $AlCl_3$ to the alcohol solution and the solution was dyed yellow.

- When adding 3-4 drops of 1% $FeCl_3$ solution to 2 ml of alcohol separation, a dark blue solution was formed.

- chromatography paper chromatogram on the chromatogram when the chromatogram is chromatographed on a chromatogram of chromatograms, chromatography paper chromatograms, chromatograms chromatography and chromatogram on 1% $AlCl_3$ from alcohol solution, with the addition of butanol fraction of the chromatography paper on the start line to the vinegar acid water (6: 4) R_f0.58; 0.68 yellow spots appear.

The above mentioned quality reactions and chromatographic analysis have shown that there are flavonoids in the composition of medicinal plants such as walnuts, peaches and MorusAlba.

3. Reaction to the coumarin compounds.

Alcoholic separation of lactone reaction from walnuts, peaches, flakes and Morus alba leafs, as well as diatoreactivity, the red color of the brown color testify to the presence of coumarins in the product.

4. Quality reactions to saponins.

- When the aqueous extract from the product is shaken hard, a constant foam has formed.

- When stable solution of 0.1 m NaOH (pH 13) with 0.1 m HCl (pH 1.0) was applied to the first of the solutions of 2 ml of aqueous dilutions, a constant foam was formed in both solutions in the same solids; but the foam level in the secondary alkaline solution was higher and stagnant (steroid saponins).

- chromatograms have a red orange color when the prepared aqueous extract is purified by chromatography and chromatogram on phosphorus hydrochloric acid at 25% solution at 105^0C for ten minutes at a temperature of 6:4. (sanonines)

5. To determine the presence of ingredients, the dry mass of crushed walnuts, peaches, vine and Morusalba leaves of 0.1 g was boiled for 10 minutes in 2-3 minutes. The mixture was cooled and filtered; Three or three drops of 1 ml of the filtrate were ironed with an ammonium acetate solution. The dark green color indicates the **presence of substances**.

Identification of flavonoids in the raw materials

The bacterial fraction of the alcohol extract from the product was chromatographed on the start line using a droplet, and the cleaner kvertsetin and routine were tested by alcohol (standard). Chromatogramis placed in a chromatographed chamber containing a mixture of acetic acid: water (6:4). After 30-40 cm rising on the system chromatography paper, chromatogram was dried, detected in UV light, spotted and treated with $AlCl_3$ 1% alcohol solution and again under UV light. Chromatogram traces were identified by R_f 0.27 kvertsetin and R_f0.64 were routinely identified by using authentic witnesses (standards).

In the chromatogram, kvertsetin and Routine Flavonoid Characteristic of Routine R_f 0.51; 0.53; 0.51; The presence of 0.61 and 0.79% was found. They are still being studied.

Determination of the sum of flavonoids. 1,0793 g peach leaf 1.0675 Morus alba leaf, 1,0258 g walnut leaf was measured at a precision cake and placed in a 25 ml measuring tube. It was filled with 3 ml of 5% alumina chloride, 70% alcohol solution, and 1 drop of diluted

acetic acid was added and 70% alcohol was added to the mark. After 40 minutes the optical density was measured at a density of 400 nm and thickness 10 mm in the cuvette.

In parallel, the optical density of the routine worker standard was measured.Flabanoid was calculated by the formula ($X_{\%}$):

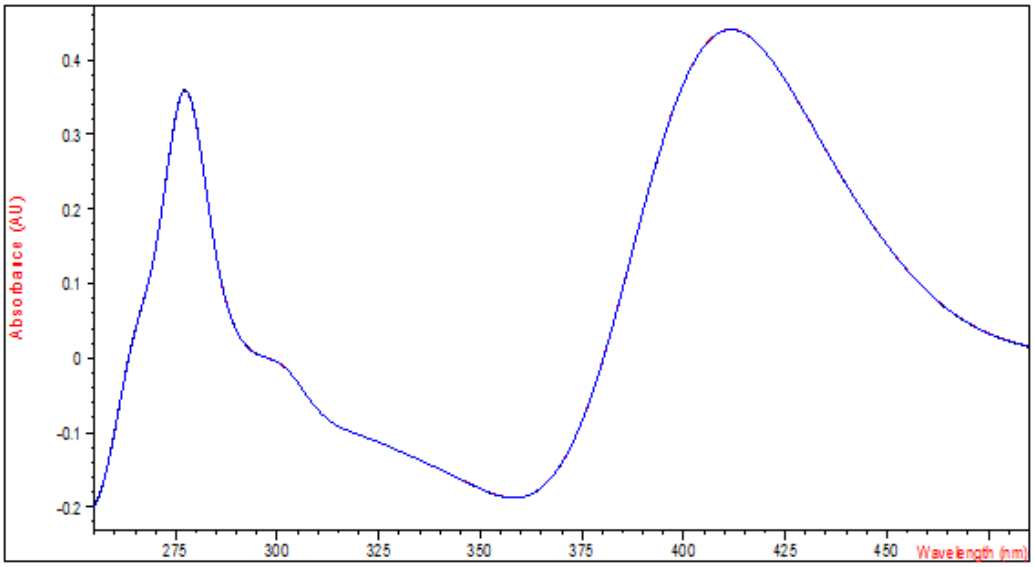
$$X = \frac{D_1 \cdot a_0 \cdot 25 \cdot 1 \cdot \mathrm{P} \cdot 100}{D_0 \cdot 3 \cdot 100 \cdot 25 \cdot 100} = \frac{D_1 \cdot a_0 \cdot \mathrm{P}}{D_0 \cdot 300};$$

D_1– optical density of the detected solution;
D_0 – optical density of the standard sample solution;
a_0 – standard specimen hood, g;
P – standard pattern follow-up routine,%.

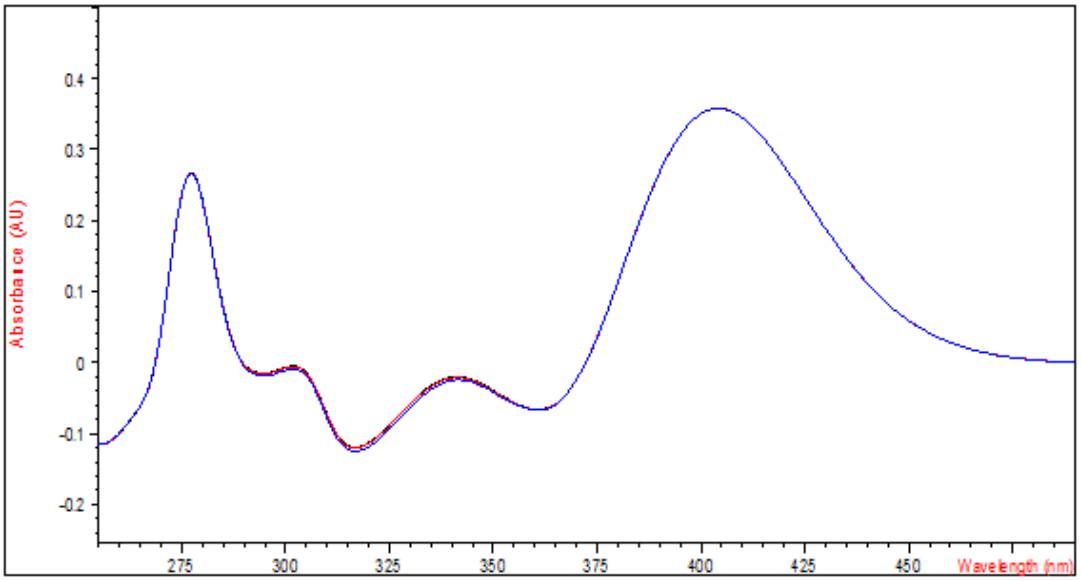
Results of the analysis and metrological characteristics of the survey are given in Table 3.

Table 3: **Determination of the amount of flavonoid in the sample**

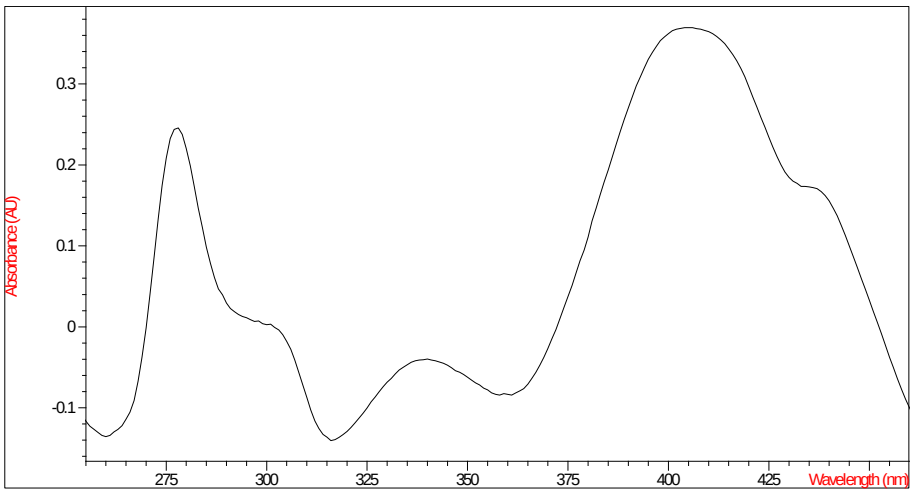
Examples	Optical density, D	Routine amount,%	Metrological characteristics
Peachleaf	0.38688 0.39211 0.39189 0.38901 0.39013	0.7561 0.7586 0.7589 0.7694 0.7784	$\bar{x}$ = 0.7643; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.00008843; S = 0.009404; $S_{\bar{X}}$ = 0.004205; ΔX= 0.02614; $\overline{\Delta X}$= 0.002614; $\mathcal{E}$ =3.42%; $\bar{\varepsilon}$ = 1.54%
Walnutleaf	0.74739 0.74648 0.74679 0.74722 0.74596	1.5232 1.5344 1.5864 1.5984 1.5746	$\bar{x}$ = 1.5634; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.00108412; S = 0.00329259; $S_{\bar{X}}$ = 0.014773; ΔX= 0.09154; $\overline{\Delta X}$= 0.00238; $\mathcal{E}$ = 0.35%; $\bar{\varepsilon}$ = 0.16%
Morus alba leaf	0.36609 0.36706 0.36334 0.36567 0.36887	0.7192 0.7245 0.7224 0.7365 0.7295	$\bar{x}$ = 0.721; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.00000368; S = 0.001918; $S_{\bar{X}}$ = 0.000858; ΔX= 0.00533; $\overline{\Delta X}$= 0.04095; $\mathcal{E}$ = 5.85%; $\bar{\varepsilon}$ = 2.62%



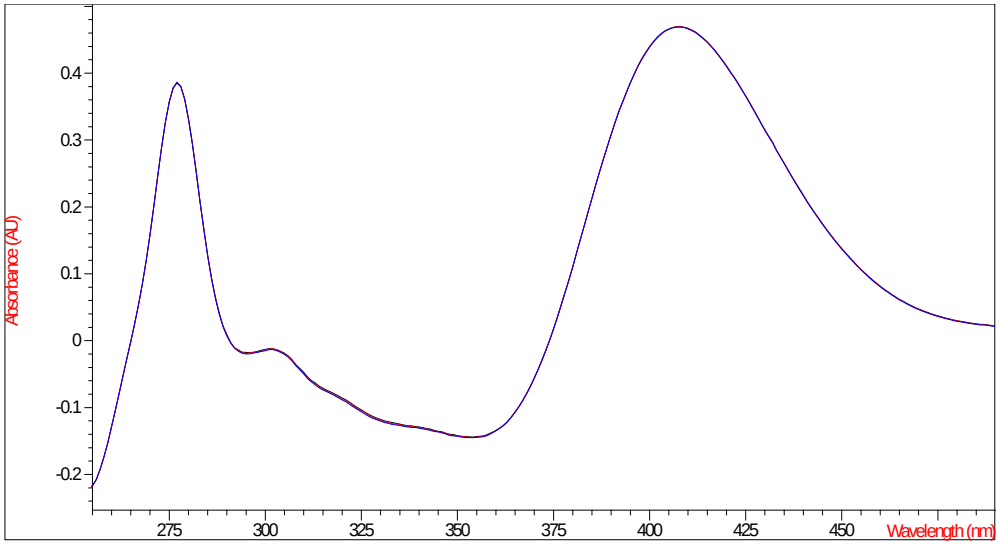
Picture 1. The UF - spectrum of the routine working standard



Picture 2. Peach leaf UF - spectrum



Picture 3. Morusalba leaf UF - spectrum



Picture 4. Walnut leaf UF - spectrum

Determination of the total flavonoids content of 40% alcohol in the morning, 40% alcohol leaf, and 70% alcohol leaf.

Extracts filtered through paper. Remove 3 ml from the filtrate and put into a 25 ml measuring tube. It was filled with 3 ml of 5% aluminum chloride in 70% alcohol solution and 1 drop of diluted acetic acid was added and 70% alcohol was added to the marker. The solution was prepared in the same manner as in the aluminum chloride solution. After 40 min, the optical density of the solution was measured at a wavelength of 400 nm and a thickness of 10 mm in the cuvette.

In parallel, the optical density of the routine worker standard was measured. Flavonoid was calculated by the formula (X%):

$$X = \frac{D_1 \cdot a_0 \cdot 25 \cdot 1 \cdot \mathrm{P} \cdot 100}{D_0 \cdot 3 \cdot 100 \cdot 25 \cdot 100} = \frac{D_1 \cdot a_0 \cdot \mathrm{P}}{D_0 \cdot 300};$$

D_1– is the optical density of the detected solution;
D_0 – optical density of the standard sample solution;
a_0 – standard specimen hood, g;
P – standard pattern follow-up routine,%.

Determination of flavonoids content of peach leaf with 70% alcohol extract, walnut leaf 70% alcohol extract, and walnut fruit 40% alcohol extracts. Filters are filtered through paper. Remove 5 ml from the filtrate and insert into a 25 ml measuring pouch and deliver 70% alcohol to the marker. The resulting solution was diluted with 3 ml and placed in a 25 ml measuring tube. It is filled with 3 ml of 5% aluminum chloride solution containing 70% alcohol solution and 1 drop of diluted acetic acid is added and 70% alcohol is added to the marker. The solution was prepared in the same manner as in the aluminum chloride solution. After 40 min, the optical density of the solution was measured at a wavelength of 400 nm and a thickness of 10 mm in the cuvette.

In parallel, the optical density of the routine worker standard was measured.

Flavonoid was dissolved by the formula (X%):

$$X = \frac{D_1 \cdot a_0 \cdot 25 \cdot 25 \cdot 1 \cdot \mathrm{P} \cdot 100}{D_0 \cdot 5 \cdot 3 \cdot 100 \cdot 25 \cdot 100} = \frac{D_1 \cdot a_0 \cdot \mathrm{P}}{D_0 \cdot 60};$$

D_1– is the optical density of the detected solution;
D_0 – optical density of the standard sample solution;
a_0 – standard specimen hood, g;
P – standard pattern follow-up routine,%.

Prepare standard sample solution. 0.05 g (net cake) Routine worker standard sample was packed into a measuring pad of 100 ml and dissolved in 70% alcohol and delivered to a mark of 70% alcohol. The resulting solution was diluted 1 ml and placed in a 25 ml measuring tube. It is filled with 1% drops of diluted acetic acid in a solution of 3 ml of 5% aluminum chloride solution containing 70% of alcohol, and 70% alcohol is added to the mark. The solution was prepared in the same manner as in the aluminum chloride solution. After 40 min, the optical density of the solution was measured at a density of 400 nm and a thickness of 10 mm in the coat (Table 4) [7, 8, 21, 22].

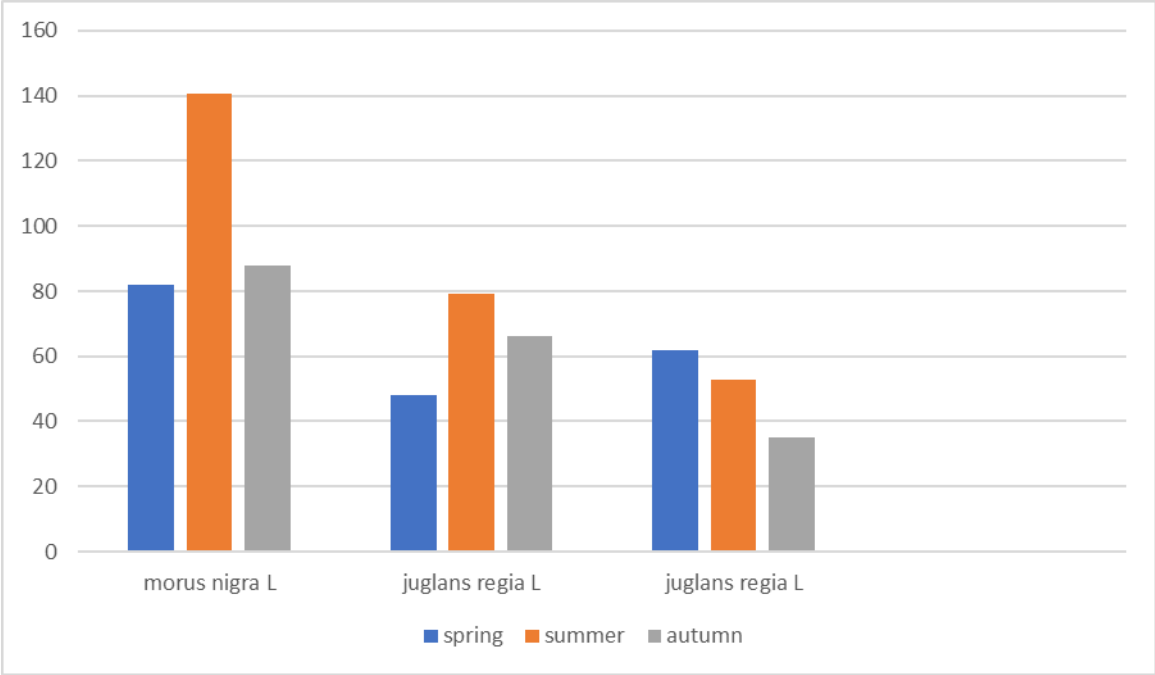
Table 4: **Determination of the amount of flavonoid in the sample**

Examples	Optical density, D	Routine amount,%	Metrological characteristics
Vine 40% alcoholcon sumption	0.12266 0.12308 0.12686 0.12476 0.12287	0.00354 0.00351 0.00355 0.00359 0.00358	$\bar{x}$ = 0.003551; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000009; S = 0.000029442; $S_{\bar{X}}$ = 0.00001317; ΔX= 0.00008185; $\overline{\Delta X}$= 0.0005660; $\mathcal{E}$ = 2.3052%; $\bar{\mathcal{E}}$ = 1.0309%
Vine leaf extract is 40% alcohol	0.34311 0.34301 0.34288 0.34301 0.34301	0.00987 0.00987 0.00986 0.00988 0.00989	$\bar{x}$ = 0.009874; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000001; S = 0.0000114018; $S_{\bar{X}}$ = 0.000005099; ΔX= 0.0000316919; $\overline{\Delta X}$= 0.0000141753; $\mathcal{E}$ = 0.32%; $\bar{\mathcal{E}}$ = 0.14%
Walnut fruit, 40% alcoholic extract	0.20405 0.20240 0.20089 0.20405 0.20605	0.00378 0.00379 0.00377 0.00379 0.00378	$\bar{x}$ = 0.003782; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000001; S = 0.00000837; $S_{\bar{X}}$ = 0.000005742; ΔX= 0.00002326; $\overline{\Delta X}$= 0.0000104; $\mathcal{E}$ = 0.62%; $\bar{\mathcal{E}}$ = 0.28%
Vineleaf 70% alcohol consumpti on	0.17371 0.17428 0.17227 0.17342 0.17342	0.00498 0.00499 0.00497 0.00498 0.00499	$\bar{x}$ = 0.004982; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000001; S = 0.00000837; $S_{\bar{X}}$ = 0.0000232591; ΔX= 0.0000104018; $\overline{\Delta X}$= 0.0000104; $\mathcal{E}$ = 0.47%; $\bar{\mathcal{E}}$ = 0.21%
Peach leaf is 70% alcoholic extract	0.35792 0.35795 0.35797 0.35785	0.05148 0.05149 0.05149 0.05149 0.05150	$\bar{x}$ = 0.004982; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000001; S = 0.00000837; $S_{\bar{X}}$ = 0.0000232591; ΔX= 0.0000104018; $\overline{\Delta X}$= 0.0000104; $\bar{x}$ $\mathcal{E}$ = 0.47%; $\bar{\mathcal{E}}$ = 0.21%
Walnut leaf 70% alcoholic extract	0.46924 0.46924 0.46950 0.46937 0.46885	0.06751 0.06752 0.06753 0.06755 0.06754	$\bar{x}$ = 0.006753; f = 4 ; T= (95%:4) = 2.78; S^2 = 0.0000000001; S = 0.000001581; $S_{\bar{X}}$ = 0.000000707; ΔX= 0.000004396; $\overline{\Delta X}$= 0.000001966; $\mathcal{E}$ = 0.065%; $\bar{\mathcal{E}}$ = 0.029%

In addition, we have studied vitamin C and common antioxidants from biologically active substances in our research. As you know, vitamins, which have high antioxidant properties, are vitamin C. In this regard, we have identified the amount of vitamin C in the spring, summer and autumn seasons from the herbaceous leaves.

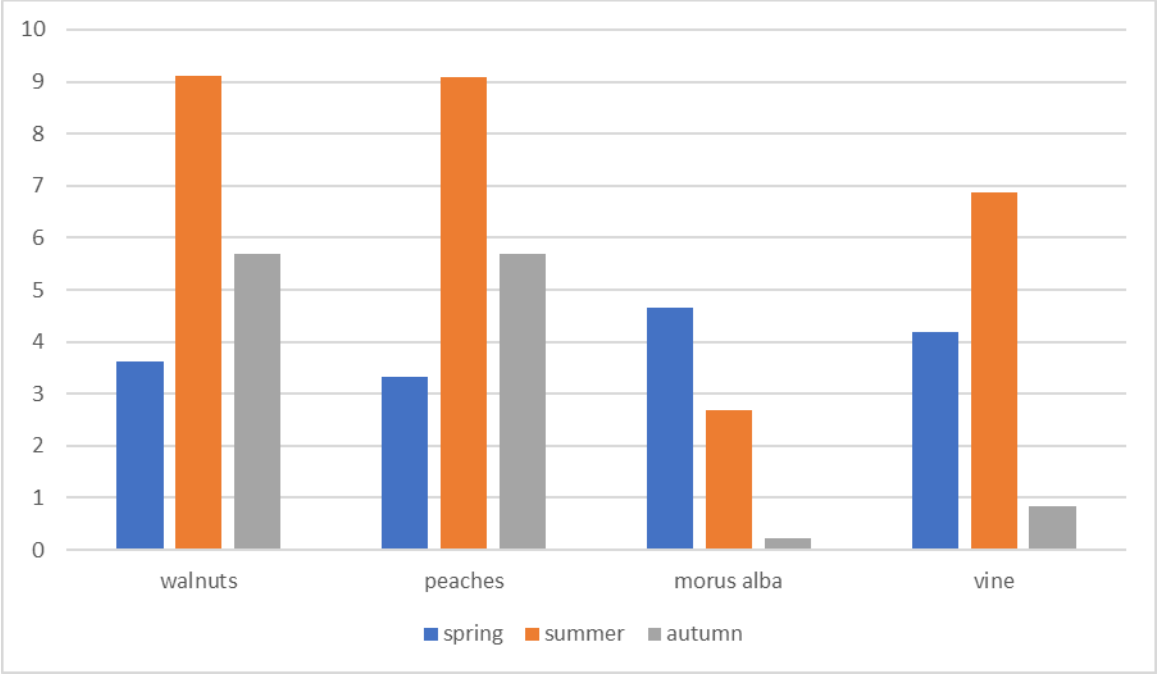
The results showed that the highest yields of plants in the shootout, peach and walnut leaves were found in different seasons: 10.8 mg% in peanut leaf, 79.2% in peach leaf, 52.8% in walnut.

Our research has shown that the amount of vitamin C in various seasons is variable, with its high concentration in the Morusalba leaf, in spring it is 81.87 mg%, in the summer - 140.8 mg% and in the autumn - 88mg (Picture 5).



Picture 5: **Vitamin C content in different seasons in plants showing antioxidant properties (mg%).**

The highest concentration of antioxidants in the summer was detected in the leaves of peaches and peaches, and its value in nut leaves was 9.105 ± 0.05 mg / g and peach leaf – 9.082 ± 0.075 mg / g. Spring walnut was 3.61 ± 0.07 mg / g and 5.7 ± 0.07 mg / g in autumn. In his fertility, the total amount of antioxidants was lower than that of the leaves. The total amount of antioxidants extracted from shutut leaf was 4.65 ± 0.55 mg / g in the spring, and 2.693 ± 0.29 mg / g in the spring, and decreased by autumn by 0.233 ± 0.07 mg / g was equal. Morusalba's boiling is used for the treatment of diabetes, skin diseases [6]. The total antioxidant content of the herbaceous plant and leaf was higher in the summer and was 6.88 ± 0.11 mg / g (Picture 6).



Picture 6: ***Total amount of antioxidants in different fruit trees (mg/g).***

The vine leaves contain vitamin C, 2% sugar, as well as carotene, kvertsetin, betaine, protokatexinate acid. In Central Asian folk medicine, young vine leaves and branches were used for the treatment of hypertension and diabetes [6].

The total amount of antioxidants in the cherry tree leaves was 6.63 ± 0.3341 mg / g, 4.632 ± 0.109 mg / g in the summer, and 3.08 ± 0.079 mg / g in the fruit.

Can be used for hepatitis treatment with fresh leaves. Boiling of all fruits, leaves and branches has anti-inflammatory, rheumatic and bleeding properties[6, 12].

Thus, in the leaves of some fruit trees showing antioxidant properties, there were common antioxidants, the highest amount of ulcers was found in the summer of walnuts and peaches.

Conclusions

Our research revealed that there are biologically active compounds in peaches, walnuts, and shrubs that show antioxidant properties in different seasons.

The findings show that there are flavonoids in medicinal plants, such as peaches, walnuts, and Morusalba leafs, and its high concentration is determined in walnut leaf.

Thus, vitamin C, flavonoids, which have antioxidant properties in medicinal herbs, can be used for correction of poisoning with pesticides from their extracts because they contain general antioxidants.

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