

ROLE OF GM-CSF IN AUTOIMMUNITY, INFLAMMATORY DISEASES, ADJUVANT TO VACCINE AND MAJOR SIDE EFFET

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ABSTRACT : Granulocyte-macrophage colony-stimulating factor (GM-CSF) is an important hematopoietic growth factor and immune modulator. During inflammation, GM-CSF can be secreted by several different cell types, including epithelial cells and leukocytes, and is a critically important cytokine that can drive both innate and adaptive immune responses. GM-CSF assembles and activates its heterodimer receptor complex on the surface of myeloid cells, initiating multiple signalling pathways that control key functions such as cell survival, cell proliferation, and functional activation. Till date, GM-CSF has been studied extensively in murine models and human clinical trials, alone and as adjuvant therapy. This knowledge provides opportunities for the development of new therapies and understands the action of these cytokines in haematological malignancy and chronic inflammation etc.

KEYWORDS : GM-CSF, Autoimmunity, Inflammatory diseases, Adjuvant.

INTRODUCTION

Granulocyte-macrophage colony-stimulating factor (GM-CSF) is an important hematopoietic growth factor and immune modulator. GM-CSF was first identified in mouse lung tissue-conditioned medium following lipopolysaccharide injection into mice by its ability to stimulate proliferation of mouse bone marrow cells *in vitro* and generate colonies of both granulocytes and macrophages. Since then, several biological effects have been attributed to GM-CSF, including hematopoiesis, response to inflammation and infection, and functional enhancement of mature effector cells in antigen presentation and cell-mediated immunity¹.

GM-CSF stimulates multipotent progenitor cells depending on its concentration, the proliferation of macrophage progenitors at the lowest doses, followed by granulocyte, erythroid, eosinophil, megakaryocyte, and multipotent progenitors². GM-CSF may play a pivotal role in various human inflammatory diseases including rheumatoid arthritis,

inflammatory renal disease, and inflammatory lung disorders. GM-CSF has been utilized in the clinical management of multiple disease processes. Most recently, GM-CSF has been incorporated into the treatment of malignancies as a sole therapy, as well as a vaccine adjuvant³. The subsequent production into a recombinant form (rHuGM-CSF; sargramostim) has led to a number of potential clinical uses of GM-CSF, including enhancement of hematopoietic recovery after chemotherapy and bone marrow transplant, treatment of infectious diseases, and use as an antitumor therapy and vaccine adjuvant⁴. So; in the present study, an attempt was made to compile the knowledge about the granulocyte-macrophage colony-stimulating factor.

GM-CSF THERAPEUTICS

A wide variety of cells can produce GM-CSF. Major sources of GM-CSF are T and B cells, monocyte/macrophage endothelial cells, and fibroblasts. Neutrophils, eosinophils, epithelial cells, mesothelial cells, Paneth cells, chondrocytes, and tumor cells can also produce GM-CSF⁵. The production of GM-CSF is stimulated by various factors, including TNF, IL-1, toll-like receptor agonists, and prostaglandin E2^{6,7}. GM-CSF promotes the survival and activation of macrophages, neutrophils, and eosinophils, as well as dendritic cell (DC) maturation. GM-CSF can polarize macrophages into M1-like inflammatory macrophages, which produce a variety of inflammatory cytokines such as TNF, IL-6, IL-12p70, IL-23, or IL-1 β , and thus promote Th1-Th17 responses^{8,9}. On the other hand, the association of GM-CSF and Th2 immunity is also reported in allergic airway inflammation¹⁰. GM-CSF is reported to have diverse functions on mature myeloid cells, including antigen presentation¹¹, induction of phagocytosis¹²⁻¹⁴, enhancement of pro-inflammatory cytokine production¹⁵, and promotion of leukocyte chemotaxis¹⁶.

GM-CSF in Auto-immunity and inflammatory diseases

Granulocyte colony-stimulating factor (G-CSF or GCSF), also known as colony-stimulating factor 3 (CSF 3), is a glycoprotein that stimulates the bone marrow to produce granulocytes and stem cells and release them into the bloodstream. In mice, transgenic studies for GM-CSF showed that many cytokines and inflammatory mediators were found to be increased in these mice and overexpression leads to macrophage accumulation, blindness, and severe damages to various tissues. GM-CSF overexpression in the stomach leads to autoimmune gastritis¹⁷⁻¹⁹. When bone marrow cells infected with a retrovirus expressing GM-CSF were transplanted, a lethal myeloproliferative syndrome was induced²⁰. In RA patients, the concentration of GM-CSF in the synovial fluid and plasma was elevated^{21,22} and the administration of recombinant GM-CSF exacerbated the disease

activity²³. Bone marrow adjacent to the RA joints contains an increased number of granulocyte-macrophage progenitors, colony-forming unit granulocyte-macrophages (CFU-GM), which can differentiate into granulocytes or macrophages with GM-CSF stimulation²⁴ and also into osteoclasts with M-CSF and RANKL stimulation²⁵. Adenoviral-mediated GM-CSF gene transfer in the lung also led to severe lung eosinophilia, macrophage expansion, and fibrotic reactions²⁶⁻²⁸. This information has led to the hypothesis that GM-CSF may have a central role in promoting sensitization to aeroallergens in polluted air²⁹⁻³⁰. Interestingly, it has been suggested that human GM-CSF polymorphisms are likely asthma determinants³¹.

In another study, it was found that the culture of ex vivo differentiated human MoDCs (CD14+CD33+) in the presence of GM-CSF is capable of class II-mediated prominent immune epitopes of two auto-antigens [type II collagen (CII) and cartilage gp39 (HCgp39)] observed in the inflamed synovial joints of patients with RA (collagen-induced arthritis (CIA), a mouse model of arthritis, mice with defective GM-CSF cannot develop arthritis, and using antibodies against GM-CSF results in inhibition of disease progression and a decrease in proinflammatory cytokines in the joints³². Similarly, in another mouse model of arthritis (in SKG mice), GM-CSF treatment increased the production of IL-1_β or IL-6 by macrophages and promoted the differentiation and augmentation of CD4⁺ T cells that produce IL-17 and GM-CSF. Also, administration of anti-GM-CSF was more efficient compared to anti-IL-17 in treatment and decreased disease severity³³.

There are four main signaling pathways triggered by CSF2R. After binding of GM-CSF to its receptor, Janus-kinase-2 (JAK-2) is recruited to the cytoplasmic domain of the β chain, and activation of JAK-2 occurs, which subsequently induces STAT-5 phosphorylation. This signaling pathway induces migration of STAT-5 dimers to the nucleus and promotes the transcription of various genes such as pim-1 and CIS to induce cell differentiation^{34,35}. GM-CSF promotes cell survival via phosphatidylinositol-3-kinase (PI3K) and JAK/STAT-Bcl-2 signaling pathways³⁶. Moreover, cell differentiation and inflammation are mediated by activation of ERK1/2 and NF-κB. Accordingly, studies have shown that GM-CSF augments the LPS-induced inflammatory response by priming of TNFα synthesis and also induces multipotent mesenteric mesothelial cell differentiation into macrophages through the ERK1/2 signaling pathway^{37,38}.

GM-CSF in crohn's disease

GM-CSF also triggers several different signalling pathways in myeloid-derived suppressor cells (MDSCs) that mainly involve the signal transducer and activator of transcription (STAT) family of transcription factors. GM-CSF is generally regarded as a cytokine with

more pro-inflammatory functions based on its activity on neutrophils and macrophages. It is likely that GM-CSF plays both protective and pathological roles³⁹. GM-CSF has been proposed to be essential for a microbiota-dependent crosstalk between mononuclear phagocytes and group 3 innate lymphoid cells (ILC3s) thereby promoting intestinal homeostasis. ILC3- derived GM-CSF can promote intestinal myeloid cell homeostasis through enhancing DC and regulatory T cell function.^{40,41}

GM-CSF-activated monocytes simultaneously have a regulatory potential on adaptive immunity. GM-CSF significantly induces a short-termed expression of chemokines in monocytes, which are known to attract naïve T cells, T helper 2 (Th2) cells, and/or regulatory T cells. GM-CSF may regulate the homing molecules CCR2 and CCR6 on human monocytes, which are involved in regulating several aspects of mucosal immunity, including the ability to mediate the recruitment of innate immune cells to the sites of epithelial inflammation⁴².

GM-CSF in Cancer

Recombinant GM-CSF has made significant contributions in the supportive care of cancer patients, owing to enhanced myeloid recovery after cytotoxic chemotherapy. GM-CSF, a potent cytokine promoting the differentiation of myeloid cells, can also be used as an immune-stimulatory adjuvant to elicit antitumor immunity⁴³. In a preclinical mouse model, the mechanism of this impairment of antitumor response by GM-CSF has also been shown to increase the generation of Foxp3⁺ Tregs and Gr-1⁺ CD11b⁺ myeloid derived suppressor cells (MDSCs) that inhibit the function of antigen-specific T cell.⁴⁴

In one of the study on tumor; it was found that antitumor growth was demonstrated in subcutaneously implanted colon cancer cell line overexpressing GM-CSF and its heterodimer receptors into immune-deficient nude mice, which exclude the interference of GM-CSF-induced immune response. The results showed significant smaller tumor burdens in-vivo in GM-CSF overexpressing compared with shRNA knockdowned GMCSF tumor cells. Moreover, the 5-year survival rate was increased in patients with colorectal cancer with concurrent overexpression of GM-CSF and its receptor subunits, suggesting a direct inhibitory role of GMCSF in tumor growth.⁴⁵

GM-CSF is a potent activation and maturation cytokine in the differentiation, maturation and migration of DCs to lymph nodes. In a murine tumor model, vaccination with GM-CSF-secreting cancer cells promotes the in vivo yields, recruitment and activation of DCs. In addition, GM-CSF stimulates an increased expression of co-stimulatory molecules B7-1 and

CD1d on DCs suggesting that a lower amount of antigen would be required to induce optimal T-cell proliferation and activation of NKT cells to execute an antitumor response.⁴⁶

GM-CSF in pregnancy

Pregnancy is associated with a transient depression of maternal cell-mediated immunity to protect the semi allogeneic embryo from rejection. Granulocyte macrophage colony-stimulating factor (GM-CSF), a lympho-haemopoietic cytokine with well-defined wide range effects covering proliferation, differentiation chemotaxis and adhesion in many cell types, plays an important role in the course of a successful pregnancy by promoting the growth and/or differentiation of the trophoblast⁴⁷. GM-CSF blood concentrations increased during normal pregnancy significantly reduced in RSA (Recurrent abortion frequency) and that such a reduction can be efficiently reverted by IVIg treatment. In fact pregnant healthy women showed very high GM-CSF blood concentrations when compared with non-pregnant healthy women. On the contrary, pregnant RSA patients did not show any increase in the GM-CSF blood concentrations, which were therefore significantly reduced in comparison with healthy pregnant women matched for age and gestational week⁴⁸.

GM-CSF in alveolar proteinosis

The absence of GM-CSF function in vivo is associated with pulmonary alveolar proteinosis (PAP), a respiratory disease characterized by an accumulation of surfactant in the lung caused by defective alveolar macrophage function.

GM-CSF, a myelopoietic growth factor and pro-inflammatory cytokine, plays a critical role in alveolar macrophage homeostasis, lung inflammation, and immunological disease. Both administration and inhibition of GM-CSF are currently being therapeutically tested in COVID-19 clinical trials. This Perspective discusses the pleiotropic biology of GM-CSF and the scientific merits behind these contrasting approaches. GM-CSF administration in patients with COVID-19 may improve lung function by strengthening the alveolar wall and enhancing viral clearance, and this approach may thus provide particular benefit in the early stages of COVID-19. By contrast, GM-CSF or GM-CSFR blockade could be a beneficial treatment for the cytokine storm and inflammatory myeloid cell tissue infiltration associated with moderate-to-severe COVID-19⁴⁹.

GM-CSF adjuvant to vaccines

GM-CSF as a sole agent in the treatment of prostate cancer and melanoma; GM-CSF has also been shown promising as a vaccine adjuvant in whole-cell, DC, and peptide-based vaccine trials for the treatment of melanoma, ovarian, colorectal, prostate, pancreatic, renal cell, and breast cancer. The efficacy of GM-CSF has been demonstrated in controlled prospective

trials, as well as in trials with comparisons to other immune adjuvants. Granulocyte-macrophage colony-stimulating factor has been used safely and effectively as a sole agent, as a byproduct of engineered tumor cells in whole-cell vaccines, as well as an adjuvant in peptide-based and DC-based vaccines used to treat multiple malignancies. Faries and colleagues reported on a randomized assessment of 97 patients with stage II-IV melanoma treated with a whole-cell allogenic vaccine (Canvaxin™; CancerVax, CA, USA) with or without GM-CSF adjuvant⁵⁰. Spitler and colleagues performed a Phase II trial investigating the use of recombinant human GM-CSF (sargramostim) in the adjuvant setting for patients with resected melanoma⁵¹. Dranoff and colleagues were able to show that vaccination with engineered GM-CSF-secreting melanoma cells augments antitumor immunity in patients with metastatic melanoma with minimal toxicities⁵².

Adjuvants like Granulocyte macrophage-colony Stimulating Factor (GM-CSF) have been found to improve the response rate to vaccines. This study was conducted to evaluate the efficacy of GM-CSF as an adjuvant to HB vaccine in ESRF patients who were non-responders to the usual three double dose vaccinations (primary non-responders). The seroconversion rate improved from an initial 62% (31/50) to an overall 84% (42/50) after the use of GM-CSF⁵³. The use of GM-CSF could augment the immunologic response to a recombinant vaccine against the hepatitis B virus (HBV) in 80 HIV-infected patients (18-35 years old). They received a double dose (40 microg) of recombinant HBV vaccine IM at 0, 1, and 6 months and were randomized to receive either concurrent 20 microg of GM-CSF (n=40) or placebo IM (n=40) with the first vaccine dose. A significant increase in the seroconversion rate was observed after the second vaccine dose in the GM-CSF group (62% GM-CSF versus 30% control group $P < 0.0074$)⁵⁴. GM-CSF as a monotherapy, as adjuvant with or without cancer vaccines, or in combination with chemotherapy.

GM-CSF SITE OF ACTION

Granulocyte-macrophage colony-stimulating factor (GM-CSF) is a pluripotent cytokine produced by many cells in the body, which regulates normal and malignant hemopoiesis as well as innate and adaptive immunity. GM-CSF assembles and activates its heterodimeric receptor complex on the surface of myeloid cells, initiating multiple signaling pathways that control key functions such as cell survival, cell proliferation, and functional activation.

GM-CSF functions through a heterodimeric receptor composed of an α -subunit that binds GM-CSF with low affinity (GMR α) and a subunit that is shared with the receptors for interleukin-3 (IL-3) and interleukin-5 (IL-5), the β common chain (bc). The bc-subunit binds cytokine very poorly by itself (McClure et al. 2003), but converts low-affinity cytokine

binding by the α -subunit to a high-affinity interaction and is the principal signal-transducing subunit. The molecular mechanisms regulating GM-CSF receptor activation have been elusive but important insights have been revealed by the recent crystal structure of the GM-CSF receptor ternary complex (Hansen et al. 2008) and are starting to clarify cytokine receptor pleiotropy as well as the function of the related IL-3 and IL-5 receptors. These

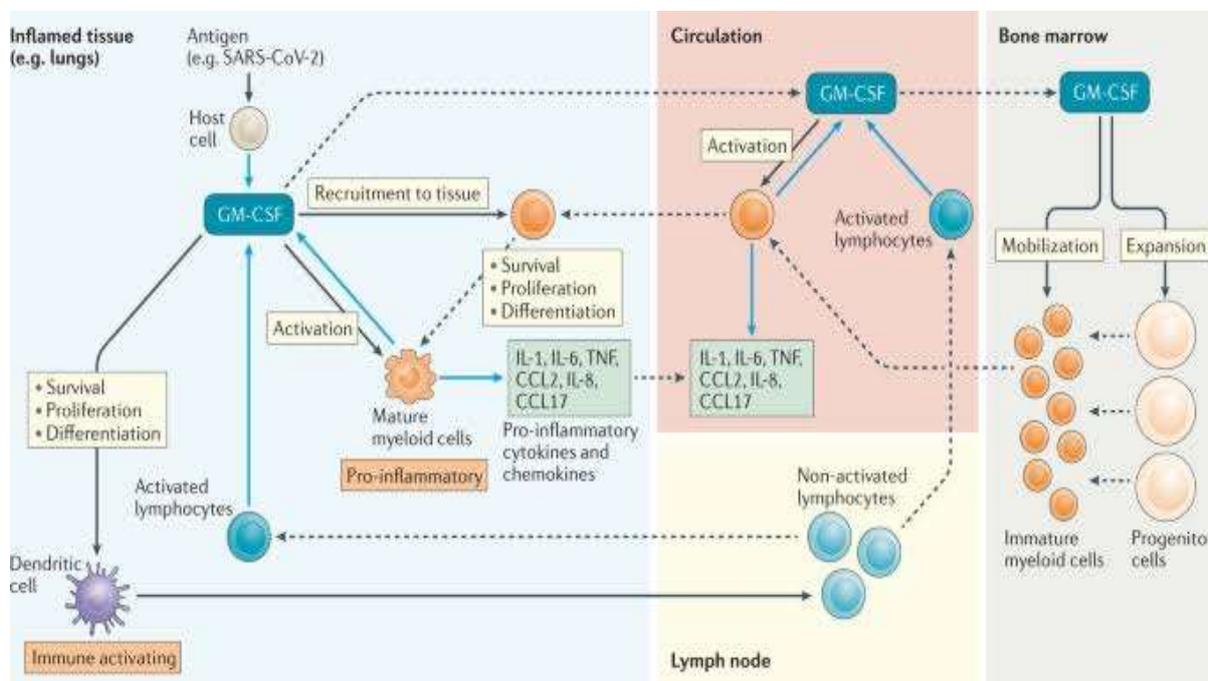


Fig. No.1 GM-CSF and Inflammation⁴⁹

insights into GM-CSF receptor activation have clinical significance and are being harnessed to develop new treatments for hematological malignancies and inflammatory diseases. Excessive stimulation of GM-CSF receptor signaling through excess and persistent GM-CSF or abnormal downstream events has been shown to contribute to chronic inflammation⁵⁵

GM-CSF has a variety of effects on the immune system including activation of T cells and maturation of dendritic cells, as well as an ability to promote humoral and cell-mediated responses. The hematopoietic cytokine granulocyte-macrophage colony-stimulating factor (GM-CSF) has been investigated as a monotherapy, and as a component of combination therapies for melanoma. Cell surface receptors for GM-CSF have been characterized by the use of radiolabeled ligand-binding studies. High-affinity GM-CSF receptors ($K_d = 30-100$ pM) are widely expressed by hematopoietic cells, with mature neutrophils expressing the highest number of receptors (500-600/cell) and immature leukemic cell lines the least (20-200 receptors/cell)^{56,57}. The mechanisms of signal transduction from the GM-CSF and IL-3 receptors are not well understood. In the neutrophil, GM-CSF enhances superoxide

production when the cells are triggered by agents, such as formyl methionyl leucine phenylalanine (fMLP) ⁵⁸. This priming does not occur directly as a result of increased calcium flux or activation of protein kinase C ⁵⁹, but is associated with enhanced Na⁺⁺in flux, release of arachidonic acid and generation of lipoxygenase products^{60,61}. An expanding role for GM-CSF in regulating immune responses has been recognized based upon its activity on the development and maturation of antigen presenting cells and its capability for skewing the immune system.

GM-CSF SIDE EFFECT

GM-CSF, have dose related side effects. GM-CSF should be used in range being from 5-10 micrograms/kg/day either by 4-6 h intravenous infusion or by subcutaneous injection. At such doses, adverse effects are predominantly mild-to-moderate in nature, occur in 20-30% of patients and usually comprise fever, myalgia, malaise, and rash or injection site reaction. Early trials using very high doses of GM-CSF were often associated with marked adverse effects, which in rare cases proved severe (pericarditis and thrombosis). Similarly, a so-called "first-dose reaction", defined as a syndrome of hypoxia and hypotension after the initial but not subsequent doses of GM-CSF, was observed in certain predisposed patients following doses above 10 micrograms/kg/day. Subsequent trials have established that intravenous bolus or short infusions of GM-CSF are more likely to promote adverse effects. Certain patient groups, for example those with myelodysplastic syndrome, acute myeloid leukaemia, inflammatory disease, autoimmune thrombocytopenia or malfunctional immunological responsiveness, require careful clinical monitoring in order to avoid potential complications following the administration of GM-CSF⁶².

CONCLUSION

Overall, GM-CSF plays an important role in inflammatory responses in autoimmune disease via induction of various cells and mediators. On-going and complete clinical trials targeting GM-CSF. Granulocyte-macrophage colony-stimulating factor (GM-CSF) can be viewed as a pro-inflammatory cytokine rather than as a key regulator of steady-state and systemic myelopoiesis. GM-CSF likely has a central role in the local activation, recruitment, and survival of macrophage lineage cells and PMNs, perhaps even contributing to macrophage proliferation a sites of inflammation. Granulocyte macrophage-colony stimulating factor (GM-CSF) is now best viewed as a major regulator governing the functions of granulocyte and macrophage lineage populations at all stages of maturation. There is recent evidence for a key role for GM-CSF in inflammatory and autoimmune diseases, therefore making it worthy

of consideration for targetting. Recently, the pathogenicity of GM-CSF-producing CD4 T cells in autoimmune and inflammatory diseases is clarified and gaining increasing attention.

REFERENCES

1. Burgess AW, Camakaris J, Metcalf D. Purification and properties of colony-stimulating factor from mouse lung-conditioned medium. *J Biol Chem.* 1977; 252:1998-2003
2. Burgess AW, Metcalf D. The nature and action of granulocyte-macrophage colony-stimulating factors. *Blood* 1980; 56: 947- 58
3. Kevin SC, Josh AT, G Travis C, Jarrod PH, Elizabeth AM, Sathibalan P et al. Use of GM-CSF as an adjuvant with cancer vaccines: beneficial or detrimental? *Expert Rev. Vaccines.* 2010; 9(5):519–525
4. Armitage JO. Emerging applications of recombinant human granulocyte-macrophage colony-stimulating factor. *Blood.* 1998; 92(12):4491–4508
5. El-Behi M, Ciric B, Dai H, Yan Y, Cullimore M, Safavi F, et al. The encephalitogenic of T(H)17 cells is dependent on IL-1- and IL-23-induced production of the cytokine GM-CSF. *Nat Immunol.* 2011; 12:568–75.
6. 4. Codarri L, Gyölvézi G, Tosevski V, Hesse L, Fontana A, Magnenat L, et al. ROR γ t drives the production of the cytokine GM-CSF in helper T cells, which is essential for the effector phase of autoimmune neuroinflammation. *Nat Immunol.* 2011; 12:560–7.
7. Verreck FA, de Boer T, Langenberg DM, Hoeve MA, Kramer M, Vaisberg E, et al. Human IL-23-producing type 1 macrophages promote but IL-10- producing type 2 macrophages subvert immunity to (myco)bacteria. *Proc Natl Acad Sci U S A.* 2004; 101:4560–5.
8. Krausgruber T, Blazek K, Smallie T, Alzabin S, Lockstone H, Sahgal N, et al. IRF5 promotes inflammatory macrophage polarization and TH1-TH17 responses. *Nat Immunol.* 2011; 12:231–8.
9. Cates EC, Fattouh R, Wattie J, InmanMD, Goncharova S, Coyle AJ, et al. Intranasal exposure of mice to house dust mite elicits allergic airway inflammation via a GM-CSF-mediated mechanism. *J Immunol.* 2004; 173:6384–92.
10. Willart MA, Deswarte K, Pouliot P, Braun H, Beyaert R, Lambrecht BN, et al. Interleukin-1 α controls allergic sensitization to inhaled house dust mite via the epithelial release of GM-CSF and IL-33. *J Exp Med.* 2012; 209:1505–17.

11. Morrissey PJ, Bressler L, Park LS, Alpert A, Gillis S. Granulocyte-macrophage colony-stimulating factor augments the primary antibody response by enhancing the function of antigen-presenting cells. *J Immunol.* 1987; 139:1113–9.
12. Berclaz PY, Zsengellér Z, Shibata Y, Otake K, Strasbaugh S, Whitsett JA, et al. Endocytic internalization of adenovirus, nonspecific phagocytosis, and cytoskeletal organization are coordinately regulated in alveolar macrophages by GM-CSF and PU.1. *J Immunol.* 2002; 169:6332–42.
13. Berclaz PY, Shibata Y, Whitsett JA, Trapnell BC. GM-CSF, via PU.1, regulates alveolar macrophage Fc γ R-mediated phagocytosis and the IL-18/IFN γ -mediated molecular connection between innate and adaptive immunity in the lung. *Blood.* 2002; 100:4193–200.
14. Fleetwood AJ, Lawrence T, Hamilton JA, Cook AD. Granulocyte-macrophage colony-stimulating factor (CSF) and macrophage CSF-dependent macrophage phenotypes display differences in cytokine profiles and transcription factor activities: implications for CSF blockade in inflammation. *J Immunol.* 2007; 178:5245–52.
15. Collins HL, Bancroft GJ. Cytokine enhancement of complement-dependent phagocytosis by macrophages: synergy of tumor necrosis factor- α and granulocyte-macrophage colony-stimulating factor for phagocytosis of *Cryptococcus neoformans*. *Eur J Immunol.* 1992; 22:1447–54.
16. Sakagami T, Uchida K, Suzuki T, Carey BC, Wood RE, Wert SE, et al. Human GM-CSF autoantibodies and reproduction of pulmonary alveolar proteinosis. *N Engl J Med.* 2009; 361:2679–81.
17. Lang RA, Metcalf D, Cuthbertson RA, et al. Transgenic mice expressing a hemopoietic growth factor gene (GM-CSF) develop accumulations of macrophages, blindness, and a fatal syndrome of tissue damage. *Cell* 1987; 51:675-86.
18. 52 Alderuccio F, Biondo M, Toh BH. Organ-specific autoimmunity in granulocyte macrophage-colony stimulating factor (GM-CSF) deficient mice. *Autoimmunity* 2002; 35:67-73.
19. 53 Biondo M, Nasa Z, Marshall A, Toh BH, Alderuccio F. Local transgenic expression of granulocyte macrophage-colony stimulating factor initiates autoimmunity. *J Immunol* 2001; 166:2090-9.
20. Johnson GR, Gonda TJ, Metcalf D, Hariharan IK, Cory S. A lethal myeloproliferative syndrome in mice transplanted with bone marrow cells infected with a retrovirus

- expressing granulocyte- macrophage colony stimulating factor. *EMBO J* 1989; 8:441-8.
21. Xu WD, Firestein GS, Taetle R, Kaushansky K, Zvaifler NJ. Cytokines in chronic inflammatory arthritis. II. Granulocyte-macrophage colony-stimulating factor in rheumatoid synovial effusions. *J Clin Invest.* 1989; 83:876–82.
 22. Fiehn C, Wermann M, Pezzutto A, Hufner M, Heilig B. Plasma GM-CSF concentrations in rheumatoid arthritis, systemic lupus erythematosus and spondyloarthritis. *Z Rheumatol.* 1992; 51:121–6.
 23. Hazenberg BP, Van Leeuwen MA, Van Rijswijk MH, Stern AC, Vellenga E. Correction of granulocytopenia in Felty's syndrome by granulocytemacrophage colony-stimulating factor. Simultaneous induction of interleukin-6 release and flare-up of the arthritis. *Blood.* 1989; 74:2769–70.
 24. Kotake S, Higaki M, Sato K, Himeno S, Morita H, Kim KJ, et al. Detection of myeloid precursors (granulocyte/macrophage colony forming units) in the bone marrow adjacent to rheumatoid arthritis joints. *J Rheumatol.* 1992; 19:1511–6.
 25. Menea C, Kurihara N, Roodman GD. CFU-GM-derived cells form osteoclasts at a very high efficiency. *Biochem Biophys Res Commun.* 2000; 267:943–6.
 26. Worgall S, Singh R, Leopold PL, *et al.* Selective expansion of alveolar macrophages in vivo by adenovirus-mediated transfer of the murine granulocyte-macrophage colony-stimulating factor cDNA. *Blood* 1999; 93:655-66.
 27. Xing Z, Ohkawara Y, Jordana M, Graham F, Gauldie J. Transfer of granulocyte-macrophage colony-stimulating factor gene to rat lung induces eosinophilia, monocytosis, and fibrotic reactions. *J Clin Invest.* 1996; 97:1102-10.
 28. Xing Z, Braciak T, Ohkawara Y, *et al.* Gene transfer for cytokine functional studies in the lung: the multifunctional role of GM-CSF in pulmonary inflammation. *J Leukoc Biol.* 1996; 59:481- 8.
 29. Frandji P, Tkaczyk C, Oskeritzian C, *et al.* Presentation of soluble antigens by mast cells: upregulation by interleukin-4 and granulocyte/ macrophage colony-stimulating factor and downregulation by interferon-gamma. *Cell Immunol.* 1995; 163:37-46
 30. Ohtoshi T, Takizawa H, Okazaki H, *et al.* Diesel exhaust particles stimulate human airway epithelial cells to produce cytokines relevant to airway inflammation in vitro. *J Allergy Clin Immunol.* 1998; 101:778-85.
 31. Delneste Y, Charbonnier P, Herbault N, *et al* Interferon-gamma switches monocyte differentiation from dendritic cells to macrophages. *Blood.* 2003; 101:143-50.

32. Cook AD, Braine EL, Campbell IK, Rich MJ, Hamilton JA. Blockade of collagen-induced arthritis post-onset by antibody to granulocytemacrophage colony-stimulating factor (GM-CSF): requirement for GMCSF in the effector phase of disease. *Arthritis Res.* 2001; 3:293–8.
33. Shiomi A, Usui T, Ishikawa Y, Shimizu M, Murakami K, Mimori T. GM-CSF but not IL-17 is critical for the development of severe interstitial lung disease in SKG mice. *J Immunol.* 2014; 193:849–59.
34. Van de Laar L, Coffey PJ, Woltman AM. Regulation of dendritic cell development by GM-CSF: molecular control and implications for immune homeostasis and therapy. *Blood.* 2012; 119:3383–93.
35. Lehtonen A, Matikainen S, Miettinen M, Julkunen I. Granulocytemacrophage colony-stimulating factor (GM-CSF)-induced STAT5 activation and target-gene expression during human monocyte/macrophage differentiation. *J Leukoc Biol.* 2002; 71:511–9.
36. Choi JK, Kim KH, Park H, Park SR, Choi BH. Granulocyte macrophage colony stimulating factor shows anti-apoptotic activity in neural progenitor cells via JAK/STAT5-Bcl-2 pathway. *Apoptosis.* 2011; 16:127–34.
37. Katz S, Zsiros V, Doczi N, Kiss AL. Inflammation-induced epithelial to mesenchymal transition and GM-CSF treatment stimulate mesenteric mesothelial cells to transdifferentiate into macrophages. *Inflammation.* 2018; 2018:825.
38. Yang TC, Chang PY, Kuo TL, Lu SC. Electronegative L5-LDL induces the production of G-CSF and GM-CSF in human macrophages through LOX-1 involving NF-kappaB and ERK2 activation. *Atherosclerosis.* 2017; 267:1–9.
39. Däbritz J. GM-CSF and the role of myeloid regulatory cells in the pathogenesis and treatment of Crohn's disease. *Molecular and Cellular Pediatrics.* 2015; 2(1):1-10
40. Sonnenberg GF, Artis D. Innate lymphoid cells in the initiation, regulation and resolution of inflammation. *Nat Med.* 2015; 21(7):698–708.
41. Eberl G, Colonna M, Di Santo JP, McKenzie AN. Innate lymphoid cells. *Innate lymphoid cells: a new paradigm in immunology. Science.* 2015; 348(6237):1-15
42. Däbritz J, Weinhage T, Varga G, Wirth T, Walscheid K, Brockhausen A, Schwarzmaier D, Bruckner M, Ross M, Bettenworth D, Roth J, Ehrchen JM, Foell D. Reprogramming of monocytes by GM-CSF contributes to regulatory immune functions during intestinal inflammation. *Journal of Immunology.* 2015; 194(5):2424–2438.

43. Yan WL, Shen KY, Tien CY, Chen YA, Liu SJ. Recent progress in GM-CSF-based cancer immunotherapy. *Immunotherapy*. 2017; 9(4):347–360
44. Jinushi M, Nakazaki Y, Dougan M, Carrasco DR, Mihm M, Dranoff G. MFG-E8-mediated uptake of apoptotic cells by APCs links the pro- and antiinflammatory activities of GMCSF. *Journal of Clinical Investigations*. 2007;117(7):1902–1913
45. Yamashita Y, Nara N, Aoki N. Antiproliferative and differentiative effect of granulocyte-macrophage colonystimulating factor on a variant human small-cell lung cancer cell line. *Cancer Res*. 1989;49(19):5334–5338
46. Mach N, Gillessen S, Wilson SB, Sheehan C, Mihm M, Dranoff G. Differences in dendritic cells stimulated in vivo by tumors engineered to secrete granulocyte-macrophage colony-stimulating factor or Flt3-ligand. *Cancer Research*. 2000;60(12), 3239–3246
47. Robertson SA, Seemark R, Guilbert LJ, Wegmann TG: The role of cytokines in gestation. *Critical Reviews on Immunology*. 1994; 14:239–292.
48. Roberto P, Caterina DC, Roberto G, Maria D, Guarino, Giuliana DS, Luigi F. GM-CSF and Pregnancy: Evidence of Significantly Reduced Blood Concentrations in Unexplained Recurrent Abortion Efficiently Reverted by Intravenous Immunoglobulin Treatment. *American Journal of Reproductive Immunology*. 2003; 50: 232–237
49. Lang FM, Lee MC, Teijaro JR et al. GM-CSF-based treatments in COVID-19: reconciling opposing therapeutic approaches. *Nat Rev Immunol*. 2020; 20: 507–514
50. Faries MB, Hsueh EC, Morton DL *et al*. Effect of granulocyte/macrophage colony-stimulating factor on vaccination with an allogenic whole-cell melanoma vaccine. *Clin. Cancer Res*. 2009; 15(22):7029–7035
51. Spitler LE, Grossbard ML, Ernstoff MS *et al*. Adjuvant therapy of stage III and IV malignant melanoma using granulocyte– macrophage colony-stimulating factor. *J Clin Oncol*. 2000; 18(8):1614–1621
52. Soiffer R, Hodi FS, Dranoff G et al. Vaccination with irradiated autologous melanoma cells engineered to secrete granulocyte–macrophage colony-stimulating factor by adenoviral-mediated gene transfer augments antitumor immunity in patients with metastatic melanoma. *J Clin Oncol*. 2003;21:3343–3350
53. Ratan Jha, Sundeep Lakhtakia, M. A. Jaleel, G. Narayan, Hemlatha K. Granulocyte-macrophage colony-stimulating factor induced sero-protection in end stage renal

- failure patients to hepatitis B in vaccine non-responders. *Renal Failure*. 2001; 23:5:629-636
54. Sasaki Md, Foccacia R, de Messias-Reason IJ. Efficacy of granulocyte-macrophage colony-stimulating factor (GM-CSF) as a vaccine adjuvant for hepatitis B virus in patients with HIV infection. *Vaccine*. 2003; 21(31):4545-9.
 55. Hercus TR, Broughton SE, Ekert PG, Ramshaw HS, Perugini M, Grimbaldston M, Lopez AF. The GM-CSF receptor family: Mechanism of activation and implications for disease. *Growth Factors*. 2012; 30(2):63–75.
 56. Gasson JC, Kaufman SE, Weisbart RH, Tomonaga M, Golde DW. High-affinity binding of granulocytemacrophage colony-stimulating factor to normal and leukemic myeloid cells. *Roc Natl Acad Sci USA* 1986;83:669-673.
 57. Park LS, Friend D, Gillis S, Urdal DL. Characterization of the cell surface receptor for granulocyte-macrophage colony-stimulating factor. *J Biol Chem* 1986;261:4177-4183.
 58. Weisbart RH, Golde DW, Clark SC, Wong GG, Gasson JC. Human granulocytemacrophage colony-stimulating factor is a neutrophil activator. *Nature* 1985;314:361-363
 59. Sullivan R, Griffin ID, Simons ER, et al. Effects of recombinant human granulocyte and macrophage colony-stimulating factors on signal transduction pathways in human granulocytes. *J Immunol* 1987;139: 3422-3430.
 60. Dahinden CA, Zing J, Maly FE, De Weck AL. Leukotriene production in human neutrophils primed by recombinant human granulocyte/macrophage colony-stimulating factor and stimulated with the complement component C5A and FMLP as second signals. *J Exp Med* 1988; 167:1281-1295.
 61. Gomez-Cambronero, Cambronero J, Yamazaki M, Mehvally F, et al. Granulocyte-macrophage colony-stimulating factor and human neutrophils: role of guanine nucleotide regulatory proteins. *Proc Natl Acad Sci USA* 1989;86:3569-3573.
 62. Stern AC, Jones TC. The side-effect profile of GM-CSF. *Infection*. 1992;20(2):S124-7