

In vitro evaluation of the anti-cancer efficacy of *Tecoma stans* hydroethanolic leaf extract against a liver cancer cell line (HepG2).

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Running Title: Anti-cancer activity of *Tecoma stans* against liver cancer cell line (HepG2)

Type of article: Original Research

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ABSTRACT

Introduction

Flavonoids, alkaloids, quinones, tannins, and traces of saponins and amino acids were found in *Tecoma stans*. The presence of phytochemicals has been recorded in almost all sections of the plant (leaves, root, flower, seed, fruit, bark) and hence the plant's usage in medical domains. Cancer, along with cardiovascular illnesses, is one of the main causes of death worldwide. The goal of this study is to see if *tecoma stans* hydro ethanolic leaf extract has anti-cancer properties against liver cancer.

Materials and methods

Human liver cancer cell line (HepG2) was brought from NCCS, Pune, India. Cell viability test and Gene expression analysis were carried out for Wnt and beta catenin mRNA Gene expression using MTT and PCR respectively. The results were analyzed using appropriate statistical tools.

Results

The Wnt and beta catenin gene expression is reduced on induction of 300 and 400 g (dosage) of hydroethanolic extract of *Tecoma stans* with significant difference in comparison with control . Thus Hydroethanolic extract of *Tecoma stans* showed significant anticancer property with the increase in dosage the anticancer activity increased contrastingly .

Conclusion

From this study the obtained results showed that *Tecoma stans* has anti cancer activity and can be used as an anti cancer drug in the medical field through the years of development of these drugs .

Key words ; *Tecoma stans* ; anti cancer ; hepatic cell line ; Wnt/beta catenin; Innovative technique.

INTRODUCTION

The plant *Tecoma stans* is a part of the family of Bignoniaceae. It is dicotyledonous in nature It has a wide range of medicinal and pharmacological applications. *Tecoma stans* is in diverse distribution in nature following the tropical and subtropical American region including Mexico to Argentina. It is also found in the Caribbean and the Bahamas. The species is characteristic of rocky slopes, often limestone outcrops , alluvial soils accompanied with drainage. It is common among the deforested and other disturbed sites and along roadways. Yellow elder has naturalized in much of tropical and subtropical Africa, Asia, the Pacific Islands and Australia (1). *Tecoma stans* is used in some parts of america as an anti anti diabetic drug. (2) .Even though *Tecoma stans* has been in medicinal use traditionally since ancient times its present implementation on smooth muscle cells remain very scarce (3) . The presence of phytochemicals has been recorded in almost all sections of the plant (leaves, root, flower, seed, fruit, bark) and hence the plant's usage in medical domains. (4) (5)

Cancer is one of the leading causes of death (6) . Normal cells have a property of contact inhibition that is during cell growth if the cell comes in contact with the cell membrane of another cell , it stops growing . Cancer cells lose the property of contact inhibition due to which they have uncontrolled cell growth and cell multiplication(4) . There are two types of cancer ; benign and malignant cancer . Benign cancer is location specific and only grown in that particular area of the body causing cancer . It can be cured by removing the tumor through surgery . The experience from our previous studies (7)

(8,9) (8)(10)(11)(12)(13)(11,13)(14)(15) (16) have led us to concentrate on the study.

Primary liver cancer, which starts in the liver, as well as malignant tumours from other regions of the body, can harm the liver. The majority of cancers are secondary or metastatic, meaning they began elsewhere in the body. Every year, cancer kills millions of people around the world. Surgery, radiation, or therapy like as chemo, hormone, and biological therapy can all be used to treat it.. Since ancient times, traditionally plants have been the source of medicines for the treatment of various diseases.(1) According to the WHO, a considerable portion of the world's population relies on plant-based medications and therapies to meet their main health-care needs.(17) Drugs derived from plants have fewer side effects compared to synthesised drugs , hence the scope for plant based medicines increases day by day . Studies at molecular levels were performed by our team of researches which insisted us to proceed this study (18–25),(26),(27),(28),(29,30),(31),(32),(33–37) The aim of this study is the evaluate the anti cancer activity of the hydroethanolic extract of leaf of *Tecoma stans* against cancer cells of liver (38)

MATERIALS AND METHODS

Cell line centre, Pune, India, provided the Human liver cancer cell line (HepG2). Tissues were cultured in RPMI media with 10% foetal bovine serum, 100 U/ml penicillin, and 100 g/ml streptomycin at 37 degrees Celsius and 5% CO₂.The MTT test was used to measure cell growth. HepG2 tissues were sown in 96-well plates with 5x10⁴/200l and grown overnight. Six duplicate wells were used in each treatment. All of the tissues were then grown for another 48 hours. The experiment was carried out three times. The MTT absorbance in negative control tissues was employed as a 0 percent cell inhibition measurement. The expression status of m RNA was analysed by Polymerase chain reaction for identifying the fold change of Wnt and beta catenin Gene expression m RNA expression over control samples. The samples were assessed using (ANOVA) and Duncan's multiple range test with p value at 0.05.

RESULT

Tecoma stans was found to reduce the abnormal proliferation of cells by reducing its cell viability at concentration (100-400 µg/ml. [Fig 1]. Effect of *tecoma stans* Wnt and Beta catenin, the cells were reduced in its proliferation and the reduction in in mRNA expression is observed. [Fig 2] [Fig 3]

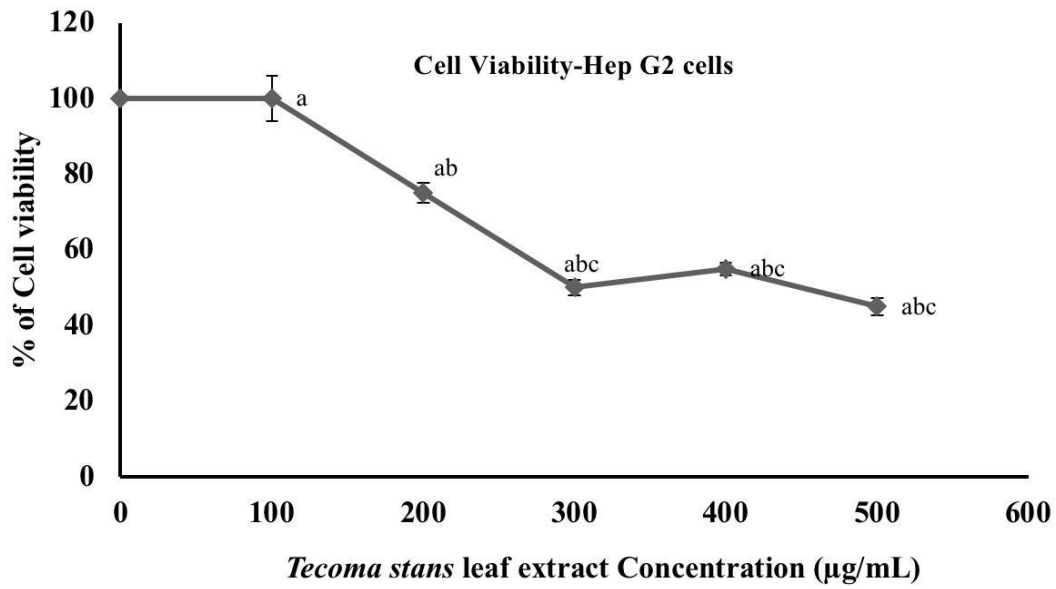


Figure 1: shows the *Tecoma stans* leaf extract on cell viability in HepG2 cells.

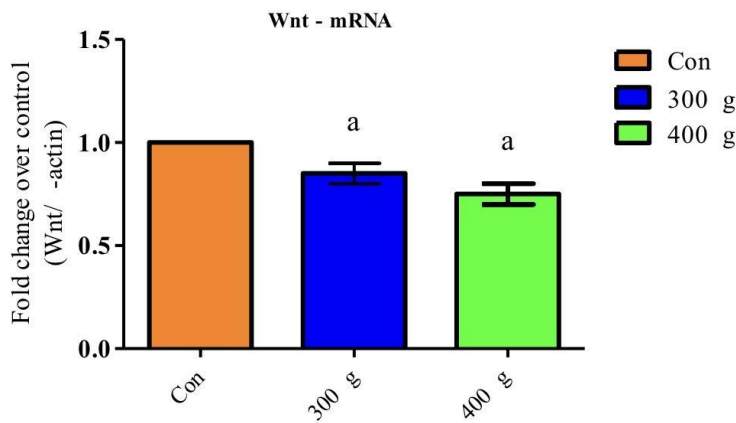


Figure:2 shows the effect of *Tecoma stans* leaf extract on Wnt mRNA expression in HepG2 cells.

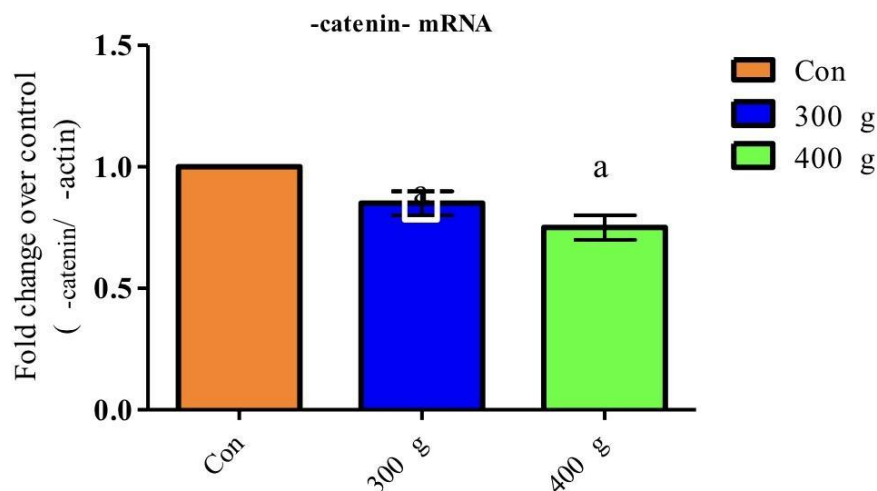


Figure :3 shows Effect of *Tecoma stans* leaf extract on B- Catenin mRNA expression in HepG2 cells

DISCUSSION

From this study it can be suggested that the Wnt and beta catenin gene expression is reduced on induction of 300 and 400 $\mu\text{g/mL}$ (dosage) of hydroethanolic extract of *Tecoma stans* with significant difference in comparison with control . Thus Hydroethanolic extract of *Tecoma stans* showed significant anticancer property with the increase in dosage the anticancer activity increased contrastingly .From this study the obtained results showed that *Tecoma stans* has anti cancer activity and can be used as an anti cancer drug in the medical field through the years of development of these drugs .

Wnt qualities encode emitted glycoproteins, which intercede intercellular flagging either over short or significant distances relying upon the tissues where they are being expressed. In creatures for the most part, Wnt flagging is engaged with a wide assortment of cell connections from early advancement through the grown-up stage. The current audit centers around the early formative jobs of Wnt qualities. The Wnt quality family, which has gone through broad quality duplications during metazoan development, can be partitioned into thirteen subfamilies, which fall into three classes varying in the sign transduction falls they trigger. Two of the flagging pathways are interrupted by Disheveled(39,40). In one, Disheveled represses GSK3, which brings about movement of β -catenin to the core. This pathway is associated with pivotal designing and detail of cell destiny. (41) This pathway intervenes planar cell extremity and concurrent augmentation developments. The third Wnt flagging pathway includes G-proteins and results in changes in intracellular Ca^{2+} levels, which assume parts in cell cycling and tissue partition.(41,42)

One of the critical components that direct cell growth, cell extremity, and cell destiny assurance during early stage improvement and tissue homeostasis is motioning by the Wnt group of secreted glycol lipoproteins.(43) Therefore, changes in the Wnt pathway are regularly connected to human birth deformities, malignant growth and different sicknesses. A basic and most contemplated Wnt pathway is authoritative Wnt flagging, what capacities by managing the measure of the transcriptional co-activator β -catenin that controls key formative quality articulation programs. This audit centers around our present comprehension of Wnt/ β -catenin flagging, drawing primarily from hereditary,

formative and biochemical examinations in *Drosophila*, *Xenopus*, mice and people. For a more exhaustive and notable viewpoint we allude to prior audits and the Wnt landing page . The nematode *Caenorhabditis elegans* shows comparative yet additionally different Wnt/ β -catenin pathways,(39) which are canvassed somewhere else and in the going with the survey . Wnt likewise initiates various non-sanctioned flagging pathways that are autonomous of β -catenin and have been as of late inspected.(44)

In tumour hosts, a decrease in viable cell count and an increase in non viable cell count toward normal suggest an anticancer effect.(45). Limitations of this study is that this is an in vitro study and future studies should be carried out invivo with large sample size to make the context evident..

CONCLUSION

From the obtained results and within limits of the study , it may be concluded that the 0hydroethanolic extract of leaf of *Tecoma stans* has a massive anti cancer potential against liver cancer by decreasing level of Wnt and beta catenin in the gene expression further studies have to be done in the future as hydroethanolic extract of leaf of *Tecoma stans* can be used as an effective anticancer drug.

ACKNOWLEDGMENT

We take this opportunity to thank Saveetha dental college and hospitals for the successful completion of the study.

Source of funding

The present study was supported by the following agencies.

- Saveetha Institute of Medical and Technical Sciences (SIMATS)
- Saveetha Dental College and Hospitals
- Saveetha University
- Dental Surgery Centre

CONFLICT OF INTEREST

All the authors declare that there was no conflict of interest in the present study .

REFERENCES

1. Abou-Alfa GK, DeMatteo R. 100 Questions & Answers About Liver Cancer. Jones & Bartlett Learning; 2019. 150 p.
2. Marzouk M, Gamal-Eldeen A, Mohamed M, El-Sayed M. Anti-proliferative and antioxidant constituents from *Tecoma stans*. *Z Naturforsch C*. 2006 Nov;61(11-12):783–91.
3. *Tecoma stans* (Linn.) H. B. & K [Internet]. SpringerReference. Available from: http://dx.doi.org/10.1007/springerreference_69544
4. Anand M, Basavaraju R. A review on phytochemistry and pharmacological uses of *Tecoma stans* (L.) Juss. ex Kunth. *J Ethnopharmacol*. 2021 Jan 30;265:113270.

5. Marzouk MSA, Gamal-Eldeen AM, Mohamed MA, El-Sayed MM. Antioxidant and Anti-Proliferative Active Constituents of *Tecoma Stans* against Tumor Cell Lines [Internet]. Vol. 1, Natural Product Communications. 2006. p. 1934578X0600100. Available from: <http://dx.doi.org/10.1177/1934578x0600100908>
6. Kameshwara S, Jothimaniv R, Senthilkum R, Kothai AR. Anti-obesity and Hypolipidemic Activity of Methanol Extract of *Tecoma stans* Flowers on Atherogenic Diet Induced Obesity in Rats [Internet]. Vol. 4, Pharmacologia. 2013. p. 77–81. Available from: <http://dx.doi.org/10.5567/pharmacologia.2013.77.81>
7. Shruthi M, Preetha S. Effect of Simple Tongue Exercises in Habitual Snorers [Internet]. Vol. 11, Research Journal of Pharmacy and Technology. 2018. p. 3614. Available from: <http://dx.doi.org/10.5958/0974-360x.2018.00665.0>
8. Preetha S, Packyanathan J. Comparison of the effect of Yoga, Zumba and Aerobics in controlling blood pressure in the Indian population [Internet]. Vol. 9, Journal of Family Medicine and Primary Care. 2020. p. 547. Available from: http://dx.doi.org/10.4103/jfmpe.jfmpe_607_19
9. J SK, Saveetha Dental College and Hospitals, Road PH, Chennai, Tamilnadu, Preetha S, et al. Effect of aerobics exercise and yoga on blood pressure in hypertensives [Internet]. Vol. 6, International Journal of Current Advanced Research. 2017. p. 3124–6. Available from: <http://dx.doi.org/10.24327/ijcar.2017.3126.0200>
10. Prathap L, Suganthirababu P, Ganesan D. Fluctuating Asymmetry of Dermatoglyphics and DNA Polymorphism in Breast Cancer Population [Internet]. Vol. 10, Indian Journal of Public Health Research & Development. 2019. p. 3574. Available from: <http://dx.doi.org/10.5958/0976-5506.2019.04141.x>
11. Lavanya J, Prathap S, Alagesan J. Digital and palmar dermal ridge patterns in population with breast carcinoma. *Biomedicine*. 2014 Jul 1;34(3):315–21.
12. Prathap L, Jagadeesan V. Association of quantitative and qualitative dermatoglyphic variable and DNA polymorphism in female breast cancer population. *Online J Health* [Internet]. 2017; Available from: https://www.researchgate.net/profile/Prathap_Suganthirababu/publication/321606278_Association_of_Quantitative_and_Qualitative_Dermatoglyphic_Variable_and_DNA_Polymorphism_in_Female_Breast_Cancer_Population/links/5a28c8f1a6fdcc8e8671c0cd/Association-of-Quantitative-and-Qualitative-Dermatoglyphic-Variable-and-DNA-Polymorphism-in-Female-Breast-Cancer-Population.pdf
13. Lavanya J, Kumar VJ, Sudhakar N, Prathap S. Analysis of DNA repair genetic polymorphism in breast cancer population. *Int J Pharma Bio Sci* [Internet]. 2015; Available from: https://scholar.google.ca/scholar?cluster=8949053652564257518&hl=en&as_sdt=0,5&sciold=0,5
14. Prathap L, Suganthirababu P. Estrogen Exposure and its Influence in DNA Repair Genetic Variants in Breast Cancer Population [Internet]. Vol. 13, Biomedical and Pharmacology Journal. 2020. p. 1321–7. Available from: <http://dx.doi.org/10.13005/bpj/2001>
15. Ravikumar H, Prathap L, Preetha S. ANALYSIS OF PALMAR ATD ANGLE IN

POPULATION WITH MALOCCLUSION. 2020 Jan 1;1174–82.

16. Prathap L. INTERPLAY OF OXIDATIVE STRESS AND LIPOPROTEINS IN BREAST CARCINOMA INITIATION, PROMOTION AND PROGRESSION -A SYSTEMATIC REVIEW. *PalArch's Journal of Archaeology of Egypt/ Egyptology* [Internet]. 2021 Jan 7 [cited 2021 Mar 9];17(7). Available from: <http://dx.doi.org/>
17. Abdeldayem H. Updates in Liver Cancer. *BoD – Books on Demand*; 2017. 222 p.
18. Sekar D, Lakshmanan G, Mani P, Biruntha M. Methylation-dependent circulating microRNA 510 in preeclampsia patients. *Hypertens Res*. 2019 Oct;42(10):1647–8.
19. Princeton B, Santhakumar P, Prathap L. Awareness on Preventive Measures taken by Health Care Professionals Attending COVID-19 Patients among Dental Students. *Eur J Dent*. 2020 Dec;14(S 01):S105–9.
20. Logeshwari R, Rama Parvathy L. Generating logistic chaotic sequence using geometric pattern to decompose and recombine the pixel values. *Multimed Tools Appl*. 2020 Aug;79(31-32):22375–88.
21. Johnson J, Lakshmanan G, M B, R M V, Kalimuthu K, Sekar D. Computational identification of MiRNA-7110 from pulmonary arterial hypertension (PAH) ESTs: a new microRNA that links diabetes and PAH. *Hypertens Res*. 2020 Apr;43(4):360–2.
22. Paramasivam A, Priyadharsini JV, Raghunandhakumar S, Elumalai P. A novel COVID-19 and its effects on cardiovascular disease. *Hypertens Res*. 2020 Jul;43(7):729–30.
23. Pujari GRS, Subramanian V, Rao SR. Effects of *Celastrus paniculatus* Willd. and *Sida cordifolia* Linn. in Kainic Acid Induced Hippocampus Damage in Rats. *Ind J Pharm Educ*. 2019 Jul 3;53(3):537–44.
24. Rajkumar KV, Lakshmanan G, Sekar D. Identification of miR-802-5p and its involvement in type 2 diabetes mellitus. *World J Diabetes*. 2020 Dec 15;11(12):567–71.
25. Ravisankar R, Jayaprakash P, Eswaran P, Mohanraj K, Vinitha G, Pichumani M. Synthesis, growth, optical and third-order nonlinear optical properties of glycine sodium nitrate single crystal for photonic device applications. *J Mater Sci: Mater Electron*. 2020 Oct;31(20):17320–31.
26. Wu S, Rajeshkumar S, Madasamy M, Mahendran V. Green synthesis of copper nanoparticles using *Cissus vitiginea* and its antioxidant and antibacterial activity against urinary tract infection pathogens. *Artif Cells Nanomed Biotechnol*. 2020 Dec;48(1):1153–8.
27. Vikneshan M, Saravanakumar R, Mangaiyarkarasi R, Rajeshkumar S, Samuel SR, Suganya M, et al. Algal biomass as a source for novel oral nano-antimicrobial agent. *Saudi J Biol Sci*. 2020 Dec;27(12):3753–8.
28. Alharbi KS, Fuloria NK, Fuloria S, Rahman SB, Al-Malki WH, Javed Shaikh MA, et al. Nuclear factor-kappa B and its role in inflammatory lung disease. *Chem Biol Interact*. 2021 Aug 25;345:109568.

29. Rao SK, Kalai Priya A, Manjunath Kamath S, Karthick P, Renganathan B, Anuraj S, et al. Unequivocal evidence of enhanced room temperature sensing properties of clad modified Nd doped mullite Bi₂Fe₄O₉ in fiber optic gas sensor [Internet]. Vol. 838, Journal of Alloys and Compounds. 2020. p. 155603. Available from: <http://dx.doi.org/10.1016/j.jallcom.2020.155603>
30. Bhavikatti SK, Karobari MI, Zainuddin SLA, Marya A, Nadaf SJ, Sawant VJ, et al. Investigating the Antioxidant and Cytocompatibility of Mimulus elengi Linn Extract over Human Gingival Fibroblast Cells. Int J Environ Res Public Health [Internet]. 2021 Jul 4;18(13). Available from: <http://dx.doi.org/10.3390/ijerph18137162>
31. Marya A, Karobari MI, Selvaraj S, Adil AH, Assiry AA, Rabaan AA, et al. Risk Perception of SARS-CoV-2 Infection and Implementation of Various Protective Measures by Dentists Across Various Countries. Int J Environ Res Public Health [Internet]. 2021 May 29;18(11). Available from: <http://dx.doi.org/10.3390/ijerph18115848>
32. Barma MD, Muthupandiyani I, Samuel SR, Amaechi BT. Inhibition of Streptococcus mutans, antioxidant property and cytotoxicity of novel nano-zinc oxide varnish. Arch Oral Biol. 2021 Jun;126:105132.
33. Vijayashree Priyadharsini J. In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens. J Periodontol. 2019 Dec;90(12):1441–8.
34. Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen A. baumannii and related species [Internet]. Vol. 94, Archives of Oral Biology. 2018. p. 93–8. Available from: <http://dx.doi.org/10.1016/j.archoralbio.2018.07.001>
35. Uma Maheswari TN, Nivedhitha MS, Ramani P. Expression profile of salivary micro RNA-21 and 31 in oral potentially malignant disorders. Braz Oral Res. 2020 Feb 10;34:e002.
36. Gudipani RK, Alam MK, Patil SR, Karobari MI. Measurement of the Maximum Occlusal Bite Force and its Relation to the Caries Spectrum of First Permanent Molars in Early Permanent Dentition. J Clin Pediatr Dent. 2020 Dec 1;44(6):423–8.
37. Chaturvedula BB, Muthukrishnan A, Bhuvanaraghan A, Sandler J, Thiruvengkatachari B. Dens invaginatus: a review and orthodontic implications. Br Dent J. 2021 Mar;230(6):345–50.
38. Curley SA. Liver Cancer. Springer Science & Business Media; 2012. 253 p.
39. Hoppler SP, Moon RT. Wnt Signaling in Development and Disease: Molecular Mechanisms and Biological Functions. John Wiley & Sons; 2014. 472 p.
40. Müschen M. WNT/ β -Catenin Signaling in Leukemia [Internet]. Targeting the Wnt Pathway in Cancer. 2011. p. 129–42. Available from: http://dx.doi.org/10.1007/978-1-4419-8023-6_6
41. Institute NC, National Cancer Institute. Wnt Family Gene Alteration Positive [Internet]. Definitions. 2020. Available from: <http://dx.doi.org/10.32388/3z85je>
42. Institute NC, National Cancer Institute. WNT Family Gene [Internet]. Definitions. 2020. Available from: <http://dx.doi.org/10.32388/6m7llc>

43. Bennett CN. Regulation of Adipocyte and Osteoblast Differentiation by Wnt Signaling. 2005.
44. Hendrix ND. The Role of WNT Signaling Pathway Defects in the Pathogenesis of Ovarian Endometrioid Adenocarcinomas. 2007.
45. Chou AH-W. The Wnt-1 Signaling Pathways in PC12 Cells. 2001. 298 p.