

Analysing Metallic Nanoparticles in Plant Extract

Yogeshwer R Ramasane¹, Sanjeev M. Reddy^{2*}

¹Research Fellow, CSIR-National Chemical Laboratory, Pune-411008, India

^{2*}Department of Chemistry, Gramin (ACS) Mahavidyalaya Vasantnagar, Mukhed Dist. Nanded (MS), India

Corresponding Author: smpreddy7@gmail.com

Abstract

Metal nanoparticles (MNPs) Medical imaging, bioengineering, photoelectricity, antibacterial, anticancer, and catalysis are just a few of the many areas where their unique physical and chemical properties have been extensively utilized. In the conventional MNP synthesis method, toxic chemicals are typically used as reducing and stabilizing agents. This method is extremely harmful to the environment and takes a long time. Because of this, environmentally friendly MNPs synthesis has recently received a lot of attention. Utilizing plant extracts as reductants and stabilizers enables MNP synthesis to be simple, cost-effective, and long-lasting. In addition, unlike their conventional counterparts, the as-synthesized MNPs are uniform in size, less toxic, and more stable. Green preparation methods are becoming an increasingly important focus in MNPs synthesis research. This systematic review provides a summary of the most recent developments in the utilization of plant extracts as reductants and stabilizers in the green synthesis of MNPs. The potential applications of MNPs made from plant extracts have also been studied in greater detail.

Keywords: *Metal NanoParticles (MPs), silver Nanoparticle (AgNP), Gold NanoParticle (AgNPs), etc*

I. Introduction

Numerous Nanotechnology has changed several research areas, such as industrial manufacturing, photoelectricity, medicine, and catalysis. In 2020, 58,000 tonnes of nanomaterials are expected to be produced industrially [1]. Metal nanoparticles (MNPs) have been extensively utilized in a number of fields, including catalysis [2, 3], electronics, optoelectronics, medicine, sensing, and information storage [4, 5] due to their distinct biological and physicochemical properties. Some of them include high electrical conductivity, high catalytic activity, high chemical stability, and biomedically important antibacterial and anticancer actions. As depicted in Figure 1, "top-down" and "bottom-up" methods are frequently employed in the synthesis of MNPs. In this method, mechanical grinding, laser ablation, and thermal decomposition, among other physical and chemical processes, are used to produce MNPs from bulk metallic materials [6].

"Bottom-up" synthesis, also known as chemical synthesis and biosynthesis, piles the metal atoms associated with MNPs [7,8]. Polyvinylpyrrolidone, formaldehyde, alkyl mercaptan, thioanthracenol, dimethylformamide, Tween 80, and formaldehyde are frequently used as stabilisers and reducing agents in chemical synthesis to produce MNPs [9]. Because hazardous chemicals are frequently utilised along this route, it is extremely detrimental for the environment. Because of the strong reduction reaction, stabilisers are frequently used when manufacturing MNPs using

conventional chemical reducing agents. The majority of the time, the costly, hazardous, and poisonous chemical reduction agents utilised in this process placed the experimenters' safety and the safety of their surroundings at jeopardy [10]. Consequently, a mild stabiliser that is also a green reducing agent with and has a mild reaction is highly desired [24- 27]

Figure 1 demonstrates that, due to their low energy consumption, low cost, and favourable environmental effects, biosynthetic techniques are substantially more environmentally friendly than chemical synthesis methods [12, 13]. However, the microbial-based synthesis is rarely utilized due to its disadvantages of being time-consuming, costly, and susceptible to biosafety concerns [14].

On the other hand, the plant-mediated synthesis of MNPs has received a lot of attention from researchers all over the world [11] due to its many benefits, such as being faster, less expensive, and easier to use with more resources. Additionally, plant extracts are friendly to the environment, inexpensive, provide MNPs with a stable protective layer, are biocompatible and biodegradable [15]. They additionally stop MNP aggregation. Nanogold, silver, copper, platinum, and palladium are just a few of the MNPs that have been the subject of recent study on the synthesis of MNPs utilising plant extracts. A brief summary of the uses of related MNPs has also been provided, which could serve as a basis and a source of ideas for future studies on the environmentally responsible synthesis of MNPs.

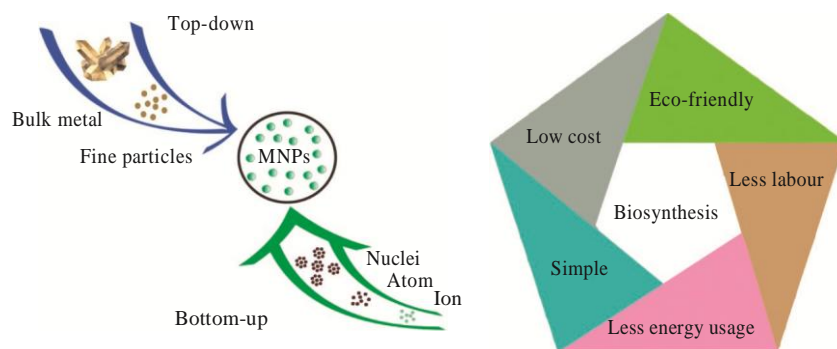


Figure 1- An Bottom up approach for Analyzing Nanoparticle in plant

II. Literature Review

1. Plant-Extract-Mediated Synthesis of Metal Nanoparticles

2.1. Synthesis of Gold Nanoparticles. Due to their stability, size controllability, biocompatibility, high adsorption capacity, high catalytic activity, and biocompatibility, gold nanoparticles (AuNPs) are a hot topic in research on nanomaterials [16]. Medicine [17], drug and gene delivery [18], biosensors [19], tomography [20], photocatalysis [21], environmental sensing [22], and water purification [23] are just a few of the many applications for AuNPs. Reducing the metal ions to metal atoms, assembling the metal atoms into nuclei through coprecipitation, sol-gel, and atomic condensation, and then growing the metal nuclei into MNPs is one of the most common "Bottom-up" synthesis techniques (Figure 1). Utilization of AuNPs in fields like biology and medicine is restricted as a result of the harmful chemicals that adsorb on their surface during their current chemical production. Consequently, green and innovative. As a result, new, environmentally friendly, and

efficient synthetic methods must immediately replace harmful chemical synthesis. It has attracted a lot of attention in this context because it is successful at creating AuNPs and because the as-prepared AuNPs are more biocompatible without containing harmful compounds [26].

By reducing chloroauric acid with ethanol extracts of powdered dried vine tea leaves, AuNPs were created (HAuCl_4). Extracts from leaves were used to create AuNPs. Additionally, they looked at the effects of reaction variables including temperature, pH, and extract dosage on the physicochemical characteristics of AuNPs. They found that vine tea extracts or alkaline circumstances caused AuNPs to aggregate excessively; smaller AuNPs could be easily synthesised at higher temperatures, whereas larger AuNPs were more stable at lower temperatures. Smaller AuNPs were easier to synthesis at higher temperatures, while larger AuNPs were more stable at lower temperatures. According to Tao and colleagues, HAuCl_4 was reduced with aloe vera leaf aqueous extracts to create spherical, highly crystalline AuNPs with particle sizes between 20 and 60 nm (28).

AuNPs were made from flower extracts; Ghosh and coworkers (29) discovered that *Gnidia glauca* flower aqueous extracts reduced HAuCl_4 and AuNPs in less than 20 minutes. There were three distinct shapes in the final AuNPs: spherical, triangular, and hexagonal, with a size of approximately 10 nm on average. The reduction of 4-nitrophenol with NaBH_4 demonstrated that the AuNPs had a significant catalytic effect on 4-aminophenol synthesis. Zangeneh and Zangeneh [30] reduced $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ to produce spherical AuNPs with a particle size range of 15-45 nm by utilizing water extracts from *Hibiscus sabdariffa* flowers. The as-synthesized AuNPs were able to significantly reduce proinflammatory cytokines while simultaneously increasing anti-inflammatory cytokines. AuNPs, like daunorubicin, did not appear to have any effect on endothelial cells.

The biological effects of the as-obtained AuNPs were examined by Baldea and colleagues along with the viability of synthesising AuNPs from aqueous extracts of *Cornusmas* fruit. Fruit or fruit shell extraction was used to create AuNPs. It was discovered that HAuCl_4 could be transformed into AuNPs at a pH of 7.5, which were fatal to hypertrophic keratinocytes. Because the AuNPs were biocompatible with common gingival fibroblasts, they might be used to treat problems related to oral dysplasia. After a 60-minute reaction at 70 °C, pH 7, 1 mM substrate, and 5 mL of fruit extracts from *Couroupita guianensis*, Sathishkumar and colleagues [31,32] produced nanogold.

The resulting AuNPs were grouped together, had a negative charge, and were coated in polyphenolic compounds. They were also negative in charge and averaged 26.11 nm in size. In addition, it was demonstrated that the as-prepared AuNPs possess exceptional hemocompatibility and anti-oxidant properties. Chen [33] produced hydrophilic AuNPs with sizes ranging from 9 to 23 nm by reducing and stabilizing them with mangosteen polyphenols from the aqueous extracts of *Garcinia mangostana* L. pericarp. He discovered that varying the extract concentration resulted in a variety of AuNP forms, including a variety of AuNP types formed.

H. Sutan [45] demonstrated that HAuCl_4 could be used to reduce ethanol extracts of *Aconitum toxicum* Reichenb roots for three hours at room temperature, resulting in the production of AuNPs with diameters ranging from 9 to 15 nm. AuNPs were produced as a result of this method. At a pH of 6.65, water extracts of *Ulva intestinalis* L. could, according to Gonzalez-Ballesteros and Associates [34], reduce chloroauric acid for four hours. At a pH of 6.65, AuNPs were made by reducing chloroauric acid for four hours. AuNPs were synthesized as a result of this. The average zeta potential and diameter of the AuNPs were 17.8 ± 2.7 nm and 22.30 ± 0.24 mV, respectively, and the antioxidant activity of the AuNPs was determined to be between 78% and 84.32%, with a total

polyphenol content of 1.49 percent and an aconitine content of 4.891 mg/mL. In a systematic investigation into their biocompatibility, AuNPs were found to be biocompatible, low in hemolytic toxicity, and capable of promoting cell proliferation.

Water, n-butanol, chloroform, and n-hexane were the four polarity solvents used to extract *Ocimum sanctum*'s active ingredient. The extracts were then used by Lee and colleagues to convert HAuCl_4 into AuNPs. They discovered that the majority of the AuNPs produced by the water extracts were thin flakes with clean edges, while the majority of the AuNPs produced by the n butanol extracts were spherical tiny particles measuring approximately 20 nm and nanosheets measuring approximately 1 μm . The majority of the AuNPs produced by the hexane extracts were nanospheres measuring less than 10 nm. The majority of the AuNPs that were produced by chloroform extracts were rough-edged nanosheets.

The green synthesis of AuNPs was made by reducing and stabilizing flavonoids, polyphenols, and amino acids from plant leaves, flowers, seeds, roots, and fruits using HAuCl_4 as a precursor (Table 1). Research on extract concentration, reaction temperature, pH, and reaction time has extensively examined AuNP morphology. Colour dye detection, antibacterial activity, and catalytic dye degradation were the most frequently utilized applications for AuNPs produced through green synthesis. The development of AuNP synthesis from plant extracts in the future looks very promising. Both the method for synthesising AuNPs on the surface of other materials through in situ reduction and research into the synthesis mechanism for establishing a precise and controllable morphology for AuNPs are crucial.

2.2. Synthesis of Silver Nanoparticles. Silver One of the nanoparticles most frequently utilized in the biomedical field is nanoparticles (AgNPs), which are an essential component of nanotechnology. AgNPs have been used in biomolecular detection, drug administration, food production, and agriculture due to their exceptional chemical stability, electrical conductivity, catalytic powers, and antibacterial properties [32]. Due to their remarkable antibacterial properties, AgNPs have emerged as one of the most promising materials for combating drug-resistant bacteria. Due to their low toxicity and biocompatibility, AgNPs made from plant extracts are better suited for use in biomedical and other fields [35].

The best reaction conditions were 60°C, 10 mL of AgNO_3 (0.005 mol/L), 60 mL of yellow quail leaf extracts (material ratio of 15 g/L), and a 40-minute reaction time when making AgNPs from *Youngia japonica* leaf extract [36]. Due to their spherical shape and average size of 20 nm, the produced AgNPs are anticipated to be used in the freshness treatment of cut flowers following the reaction, which effectively reduced the bacteria that were detached from cut lilies' stem ends. By reducing silver nitrate with water-soluble green tea extracts, Wang and his colleagues (37) produced spherical AgNPs with a range of particle sizes between 30 and 40 nm.

Table 1- Green synthesis of silver nanoparticles by different researchers using plant extracts.

Plants	Size (nm)	Plant's part	Shape
--------	-----------	--------------	-------

<i>Alternanthera dentate</i>	50–85	Leaves	Spherical
<i>Acorus calamus</i>	31.86	Rhizome	Spherical
<i>Boerhaavia diffusa</i>	24	Whole plant	Spherical
Tea extract	20–80	Leaves	Spherical
<i>Tribulus terrestris</i>	16–27	Fruit	Spherical
<i>Cocous</i>	23	Inflorescence	Spherical
<i>Abutilon indicum</i>	8–17	Leaves	Spherical
<i>Pistacia atlantica</i>	11–50	Seeds	Spherical
<i>Ziziphora tenuior</i>	9–40	Leaf	Spherical
<i>Ficus carica</i>	14	Leaf	–
<i>Cymbopogan citratus</i>	33	Leaf	–
<i>Acalypha indica</i>	0.7	Leaf	–
<i>Premna herbacea</i>	11–30	Leaf	Spherical
<i>Calotropis procera</i>	18–45	Plant	Spherical
<i>Centella asiatica</i>	32–50	Leaf	Spherical
<i>Argyreia nervosa</i>	22–50	Seed	–
<i>Psoralea corylifolia</i>	80–110	Seed	–
<i>Brassica rapa</i>	17.4	Leaf	–
<i>Coccinia indica</i>	11–20	Leaf	–
<i>Vitex negundo</i>	5 & 10–30	Leaf	Spherical & fcc
<i>Melia dubia</i>	35	Leaf	Spherical
<i>Portulaca oleracea</i>	<60	Leaf	–

III Antimicrobial property of silver nanoparticles and its mechanism

All civilizations have made substantial use of silver for a number of purposes. Fine cutlery, decorations, and jewellery are all made out of silver in various societies. It was believed that wearing silver jewellery, handicrafts, and silverware was healthy. Silver has been utilised as an antibacterial since the Phoenicians employed it as a natural biocide to protect milk bottles against microorganism infection. 650 distinct types of microorganisms, including viruses, fungi, gram-positive and gram-negative bacteria, and others, can all be killed by the well-known antibiotic silver. An application for the metal that is still relatively new is in silver nanoparticles. Silver has been mentioned as a therapy for several illnesses in the traditional Ayurvedic medicine of ancient India. Applying drops of aqueous silver nitrate to new borns' eyes during childbirth to stop the spread of *Neisseria gonorrhoea* from infected moms became standard procedure [41-43]. Silver has been discovered to be the least hazardous to animal cells and the most effective antibacterial agent of all the metals possessing antimicrobial capabilities. Since more than 2,000 years ago, silver nitrate has been used to treat infections. However, when silver nanoparticles are used, the surface area to which microbes can be exposed increases significantly. For instance, during the World War, silver was frequently employed in the medical care of injured soldiers. Numerous plant extracts from different sources have been examined for their antibacterial capabilities. Plant extracts from a variety of sources have been used in analyses of the antimicrobial properties of silver nanoparticles against various microbes [38, 39]

The antimicrobial properties of silver nanoparticles depend on:

1. Capping agent and environmental conditions (size, pH, and size).

2. The precise antibacterial or toxicological processes of silver nanoparticles are still the subject of extensive research and discussion.

. The positive charge of the Ag ions is thought to be essential for the antibacterial properties. Silver must be ionised in order to possess any antibacterial effects. Silver is inert while it is not ionised, but when it comes into contact with moisture, silver ions are released. Ag⁺ ions can form complexes with nucleic acids and prefer to bind with nucleosides over phosphate groups. All silver-containing compounds and silver-containing compounds with known antibacterial activities result in the formation of silver ions (Ag⁺).

These silver ions can be produced by ionizing the surface of a solid silver particle in the case of silver nanoparticles. They are also able to be incorporated into silver sulfadiazine and released gradually over time. Positively charged nanoparticles and negatively charged bacterial cells electrostatically attract each other, making them the ideal bactericide, according to some published studies.

It has been demonstrated that these nanoparticles accumulate within the membrane before entering the cell and causing damage to the membrane or cell wall. Silver atoms are thought to bind to enzyme thiol groups (ASH) and form stable SAAg bonds with thiol-containing molecules, deactivating enzymes that transport ions and generate trans membrane energy. As a result of the breakdown of the hydrogen bonding between the two anti-parallel strands, it was anticipated that the DNA molecule would denaturate once the Ag(I) ion entered the cell and intercalated between the purine and pyrimidine base pairs(40)

Bacterial cell lysis may account for its antibacterial properties. Nanoparticles altered the phosphotyrosine profile of the bacterial peptide, which interfered with signal transduction and prevented microbial growth. The emergence of bacteria that are resistant to antibiotics has no impact on the antibacterial action, which is dose-dependent. Following treatment, it was discovered that silver nanoparticles accumulated in the bacterial membrane of E. coli cells, causing the cell to die and become more permeable.

Conclusion

The utilisation of green chemistry and a green approach to create metal nanoparticles has increased interest in creating ecologically friendly processes. The production of silver nanoparticles using plant extracts has the benefit of being economical, cost-effective, and energy-efficient. It also makes communities and workplaces healthier while simultaneously preserving consumer safety, environmental protection, and human health. Silver nanoparticles made using green synthesis have significant nanotechnology features that can be applied in ways that are unmatched by other materials. Silver nanoparticles made using green synthesis have significant nanotechnology features that can be applied in ways that are unmatched by other materials. Due to the time-consuming process of using microorganisms and maintaining their culture, which can limit their potential for the production of nanoparticles, plants may be a better choice than other biological entities for this purpose. The use of plant extract in synthesis may therefore have a significant impact in the future.

The manufacture of silver nanoparticles utilising plant extracts, such the ones mentioned before, has been the subject of numerous reports. There hasn't been any research on how to economically and environmentally uncover the natural reducing constituent's capacity to make silver nanoparticles. Lab results may vary because plant extracts from the same species gathered in various parts of the world have chemical compositions that range greatly from one another. When employing

plant extracts as reducing and stabilising agents to make silver nanoparticles, this is the main problem that appears, and it needs to be fixed. A simple and quick fix for the aforementioned issue could be provided by identifying the plant biomolecules that mediate the creation of nanoparticles give green syntheses of silver nanoparticles a new lease on life.

Reference

- [1] R. A. Kudgus, K. Giri, R. Bhattacharya et al., "Intrinsic therapeutic applications of noble metal nanoparticles: past, present and future," *Chemical Society Reviews*, vol. 41, no. 7, pp. 2943–2970, 2012.
- [2] S. E. Skrabalak, L. Au, X. Lu, X. Li, and Y. Xia, "Gold nanocages for cancer detection and treatment," *Nanomedicine*, vol. 2, no. 5, pp. 657–668, 2007.
- [3] E. Antolini, "Palladium in fuel cell catalysis," *Energy & Environmental Science*, vol. 2, no. 9, pp. 915–931, 2009.
- [4] F. T. Minhas, G. Arslan, I. H. Gubbuk et al., "Evaluation of antibacterial properties on polysulfone composite membranes using synthesized biogenic silver nanoparticles with *Ulva compressa* (L.) Kutz." and *Cladophora glomerata* (L.) Kutz". extracts," *International Journal of Biological Macromolecules*, vol. 107, pp. 157–165, 2018.
- [5] P. Dauthal and M. Mukhopadhyay, "Noble metal nanoparticles: plant-mediated synthesis, mechanistic aspects of synthesis, and applications," *Industrial & Engineering Chemistry Research*, vol. 55, no. 36, pp. 9557–9577, 2016.
- [6] M. A. Meyers, A. Mishra, and D. J. Benson, "Mechanical properties of nanocrystalline materials," *Progress in Materials Science*, vol. 51, no. 4, pp. 427–556, 2006.
- [7] K. N. Thakkar, S. S. Mhatre, and R. Y. Parikh, "Biological synthesis of metallic nanoparticles," *Nanomedicine: Nanotechnology, Biology and Medicine*, vol. 6, no. 2, pp. 257–262, 2010.
- [8] P. Mukherjee, A. Ahmad, D. Mandal et al., "Fungusmediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: a novel biological approach to nanoparticle synthesis," *Nano Letters*, vol. 1, no. 10, pp. 515–519, 2001.
- [9] H. Lee, A. K. R. Lytton-Jean, Y. Chen et al., "Molecularly selfassembled nucleic acid nanoparticles for targeted in vivo siRNA delivery," *Nature Nanotechnology*, vol. 7, no. 6, pp. 389–393, 2012.
- [10] D. Nath and P. Banerjee, "Green nanotechnology-a new hope for medical biology," *Environmental Toxicology and Pharmacology*, vol. 36, no. 3, pp. 997–1014, 2013.
- [11] K. B. Narayanan and N. Sakthivel, "Green synthesis of biogenic metal nanoparticles by terrestrial and aquatic phototrophic and heterotrophic eukaryotes and biocompatible agents," *Advances in Colloid and Interface Science*, vol. 169, no. 2, pp. 59–79, 2011.
- [12] P. Raveendran, J. Fu, and S. L. Wallen, "Completely "green" synthesis and stabilization of metal nanoparticles," *Journal of the American Chemical Society*, vol. 125, no. 46, pp. 13940-13941, 2003.
- [13] S. A. Akintelu, S. C. Olugbeko, F. A. Folorunso, A. K. Oyebamiji, and A. S. Folorunso, "Characterization and pharmacological efficacy of silver nanoparticles biosynthesized using the bark extract of *Garcinia Kola*," *Journal of Chemistry*, vol. 2020, pp. 2876019–2876025, 2020.
- [14] V. Kumar and S. K. Yadav, "Plant-mediated synthesis of silver and gold nanoparticles and their applications," *Journal of Chemical Technology & Biotechnology*, vol. 84, no. 2, pp. 151–157, 2009.
- [15] X. Wang, L. Yuan, H. Deng, and Z. Zhang, "Structural characterization and stability study of green synthesized starch stabilized silver nanoparticles loaded with isoorientin," *Food Chemistry*, vol. 338, pp. 127807–127809, 2021.
- [16] R. Guo, Y. Song, G. Wang, and R. W. Murray, "Does core size matter in the kinetics of ligand exchanges of monolayerprotected Au clusters?" *Journal of the American Chemical Society*, vol. 127, no. 8, pp. 2752–2757, 2005.

- [17] D. Pissuwan, C. H. Cortie, S. M. Valenzuela, and M. B. Cortie, "Functionalised gold nanoparticles for controlling pathogenic bacteria," *Trends in Biotechnology*, vol. 28, no. 4, pp. 207–213, 2010.
- [18] P. Ghosh, G. Han, M. De, C. Kim, and V. Rotello, "Gold nanoparticles in delivery applications," *Advanced Drug Delivery Reviews*, vol. 60, no. 11, pp. 1307–1315, 2008.
- [19] S. Singh, D. V. S. Jain, and M. L. Singla, "Sol-gel based composite of gold nanoparticles as matrix for tyrosinase for amperometric catechol biosensor," *Sensors and Actuators B: Chemical*, vol. 182, pp. 161–169, 2013.
- [20] H. Liu, Y. Xu, S. Wen et al., "Facile hydrothermal synthesis of low generation dendrimer-stabilized gold nanoparticles for in vivo computed tomography imaging applications," *Polymer Chemistry*, vol. 4, no. 6, pp. 1788–1795, 2013.
- [21] B. Paul, B. Bhuyan, D. Dhar Purkayastha, M. Dey, and S. S. Dhar, "Green synthesis of gold nanoparticles using *Pogostemon benghalensis* (B) O. Ktz. leaf extract and studies of their photocatalytic activity in degradation of methylene blue," *Materials Letters*, vol. 148, pp. 37–40, 2015.
- [22] K. Saha, S. S. Agasti, C. Kim, X. Li, and V. M. Rotello, "Gold nanoparticles in chemical and biological sensing," *Chemical Reviews*, vol. 112, no. 5, pp. 2739–2779, 2012.
- [23] T. Pradeep and Anshup, "Noble metal nanoparticles for water purification: a critical review," *Thin Solid Films*, vol. 517, no. 24, pp. 6441–6478, 2009.
- [24] M. Yadi, E. Mostafavi, B. Saleh et al., "Current developments in green synthesis of metallic nanoparticles using plant extracts: a review," *Artificial cells, nanomedicine, and biotechnology*, vol. 46, pp. S336–S343, 2018.
- [25] L. Castillo-Henríquez, K. Alfaro-Aguilar, J. Ugalde-Alvarez, L. Vega-Fernandez, G. Montes de Oca-Vasquez, and J. R. Vega-Baudrit, "Green synthesis of gold and silver nanoparticles from plant extracts and their possible applications as antimicrobial agents in the agricultural area," *Nanomaterials*, vol. 10, no. 9, pp. 1763–1786, 2020.
- [26] S. K. Nune, N. Chanda, R. Shukla et al., "Green nanotechnology from tea: phytochemicals in tea as building blocks for production of biocompatible gold nanoparticles," *Journal of Materials Chemistry*, vol. 19, no. 19, pp. 2912–2920, 2009.
- [27] Q. Guo, Z. Fu, C. Dong et al., "Biosynthesis of gold nanoparticles using vine tea powder extracts," *Chinese Journal of Applied Chemistry*, vol. 31, no. 7, pp. 841–846, 2014.
- [28] J. Tao, Z. Fu, C. Dong, X. Wang, and X. Yang, "Green synthesis and characterization of monodisperse gold nanoparticles using aloe vera leaf extract," *Rare Metal Materials and Engineering*, vol. 48, no. 11, pp. 3470–3475, 2019.
- [29] S. Ghosh, S. Patil, M. Ahire et al., "*Gnidia glauca* flower extract mediated synthesis of gold nanoparticles and evaluation of its chemocatalytic potential," *Journal of Nanobiotechnology*, vol. 10, no. 1, pp. 17–25, 2012.
- [30] M. M. Zangeneh and A. Zangeneh, "Novel green synthesis of *Hibiscus sabdariffa* flower extract conjugated gold nanoparticles with excellent anti-acute myeloid leukemia effect in comparison to daunorubicin in a leukemic rodent model," *Applied Organometallic Chemistry*, vol. 34, no. 1, pp. 5271–5283, 2020.
- [31] I. Baldea, A. Florea, D. Olteanu et al., "Effects of silver and gold nanoparticles phytosynthesized with *Cornus mas* extract on oral dysplastic human cells," *Nanomedicine*, vol. 15, no. 1, pp. 55–75, 2020.
- [32] G. Sathishkumar, P. K. Jha, V. Vignesh et al., "Cannonball fruit (*Couroupita guianensis*, Aubl.) extract mediated synthesis of gold nanoparticles and evaluation of its antioxidant activity," *Journal of Molecular Liquids*, vol. 215, pp. 229–236, 2016.
- [33] L. Chen, "Study on biosynthesis and spectral property of gold nanoparticles in the extracts of mangosteen (*Garcinia mangostana* L) pericarp," *Chemical Research and Application*, vol. 26, no. 1, pp. 74–80, 2014.
- [34] S. Patra, S. Mukherjee, A. K. Barui, A. Ganguly, B. Sreedhar, and C. R. Patra, "Green synthesis, characterization of gold and silver nanoparticles and their potential application for cancer therapeutics," *Materials Science and Engineering: C*, vol. 53, pp. 298–309, 2015.

- [35] S. S. Shankar, A. Ahmad, R. Pasricha, and M. Sastry, "Bioreduction of chloroaurate ions by geranium leaves and its endophytic fungus yields gold nanoparticles of different shapes," *Journal of Materials Chemistry*, vol. 13, no. 7, pp. 1822–1826, 2003.
- [36] N. U. Islam, K. Jalil, M. Shahid et al., "Green synthesis and biological activities of gold nanoparticles functionalized with *Salix alba*," *Arabian Journal of Chemistry*, vol. 12, no. 8, pp. 2914–2925, 2019.
- [37] V. Karthika, A. Arumugam, K. Gopinath et al., "Guazuma ulmifolia bark-synthesized Ag, Au and Ag/Au alloy nanoparticles: photocatalytic potential, DNA/protein interactions, anticancer activity and toxicity against 14 species of microbial pathogens," *Journal of Photochemistry and Photobiology B: Biology*, vol. 167, pp. 189–199, 2017.
- [38] R. Majumdar, B. G. Bag, and P. Ghosh, "Mimusops elengi bark extract mediated green synthesis of gold nanoparticles and study of its catalytic activity," *Applied Nanoscience*, vol. 6, no. 4, pp. 521–528, 2016.
- [39] A. C. Barai, K. Paul, A. Dey et al., "Green synthesis of Nerium oleander-conjugated gold nanoparticles and study of its in vitro anticancer activity on MCF-7 cell lines and catalytic activity," *Nano Convergence*, vol. 5, no. 1, pp. 10–18, 2018.
- [40] A. K. Singh, Y. B. Tripathi, N. Pandey, D. P. Singh, D. Tripathi, and O. N. Srivastava, "Enhanced antilipopopolysaccharide (LPS) induced changes in macrophage functions by *Rubia cordifolia* (RC) embedded with Au nanoparticles," *Free Radical Biology and Medicine*, vol. 65, pp. 217–223, 2013.
- [41] D. Van-Dat, T. Anh Tai, N. Thanh-Danh et al., "Biosynthesis of gold nanoparticles using *Litsea cubeba* fruit extract for catalytic reduction of 4-nitrophenol," *Journal of Nanomaterials*, vol. 2020, Article ID 4548790, 10 pages, 2020.
- [42] J. R. Nakkala, R. Mata, and S. R. Sadras, "The antioxidant and catalytic activities of green synthesized gold nanoparticles from *Piper longum* fruit extract," *Process Safety and Environmental Protection*, vol. 100, pp. 288–294, 2016.
- [43] S. Naraginti, P. L. Kumari, R. K. Das, A. Sivakumar, S. H. Patil, and V. V. Andhalkar, "Amelioration of excision wounds by topical application of green synthesized, formulated silver and gold nanoparticles in albino Wistar rats," *Materials Science and Engineering: C*, vol. 62, pp. 293–300, 2016.
- [44] P. Khademi-Azandehi and J. Moghaddam, "Green synthesis, characterization and physiological stability of gold nanoparticles from *Stachys lavandulifolia* Vahl extract," *Particuology*, vol. 19, pp. 22–26, 2015.
- [45] N. A. Sutan, D. S. Manolescu, I. Fierascu et al., "Phytosynthesis of gold and silver nanoparticles enhance in vitro antioxidant and mitostimulatory activity of *Aconitum toxicum* Reichenb. rhizomes alcoholic extracts," *Materials Science and Engineering: C*, vol. 93, pp. 746–758, 2018.